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Multi-plug Filtration Clean-up (m-PFC) Method and Automated Device for Analysis of Pesticide and Veterinary Drug Residues

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China Agricultural University

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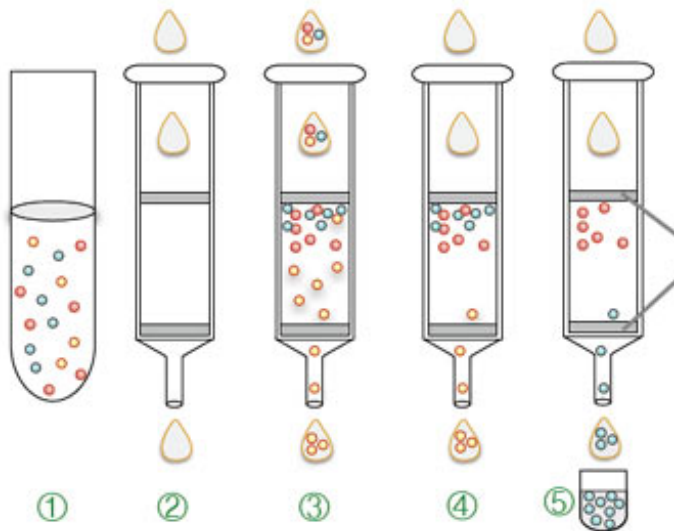
I. d-SPE method and **optimization** based on nano-materials cleanup

II. **m-PFC** method development and application

III. m-PFC **automatic** device and evaluation

Summary & Discussion

Pesticide Residue Cleanup Methods



I. d-SPE method based on Carbon Nanotubes -Optimization on QuEChERS method

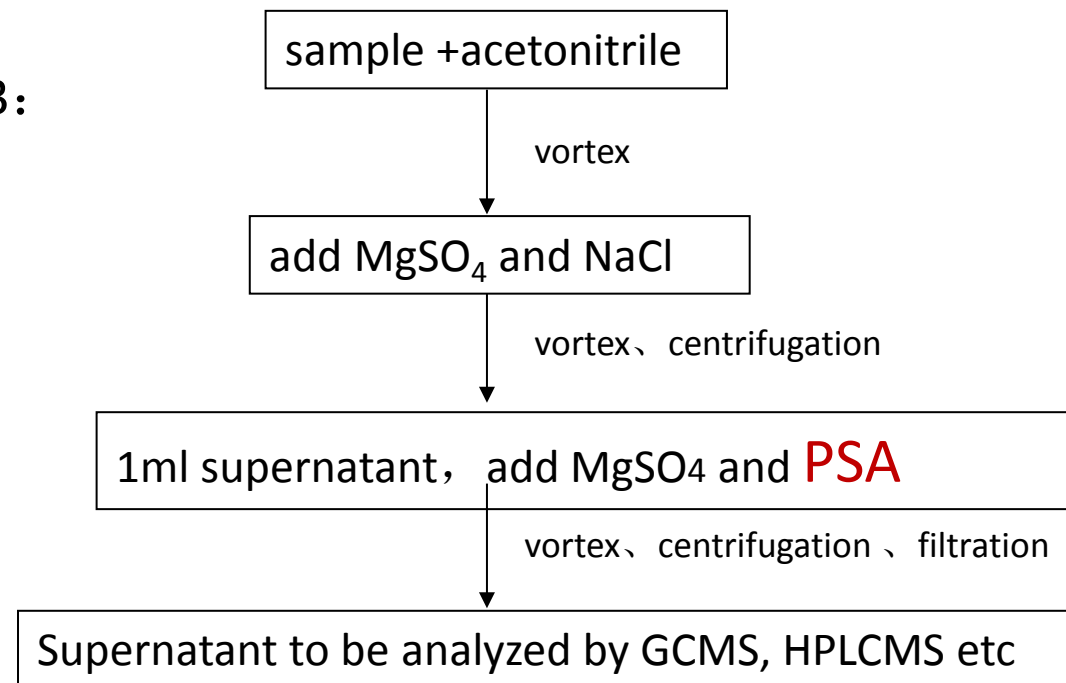
QuEChERS: by Steve Lehotay, Anastassiades et al. since 2003:

from SPE to r-DSE

Methods: AOAC: 2007.01

EU: EN15662

China: GB 23200.113-2018 etc.



Advantage: Quick

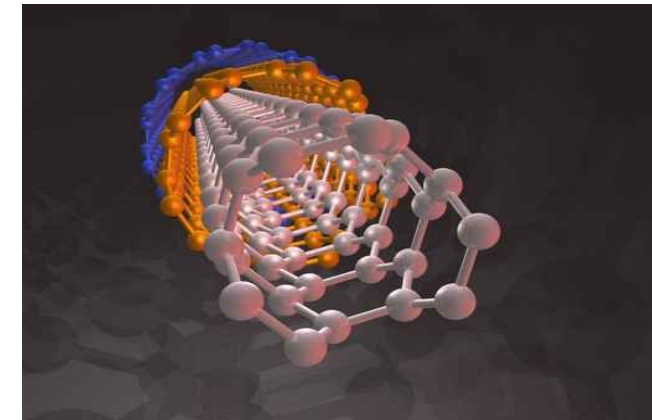
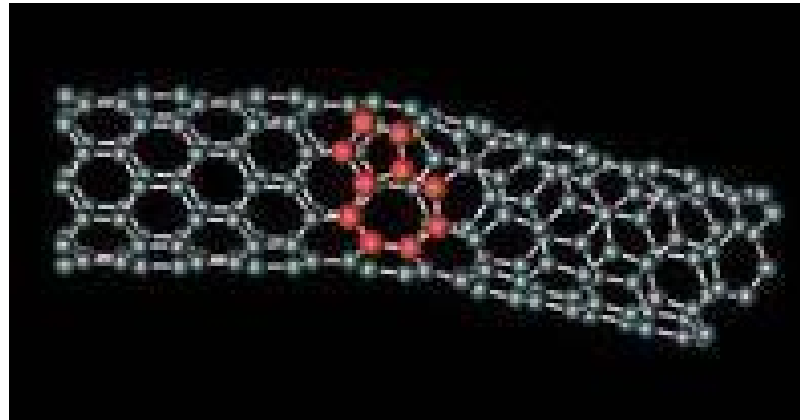
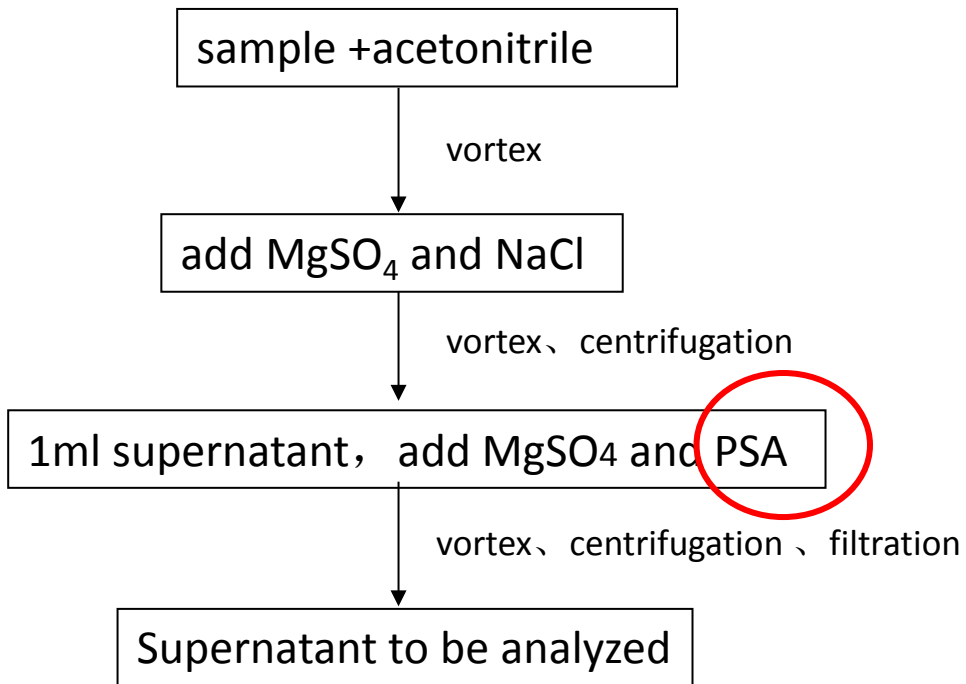
Disadvantage: cleanup pe



Which is ready for Injection ?

I. d-SPE method optimization based on MWCNTs

Finding other absorbents and combinations

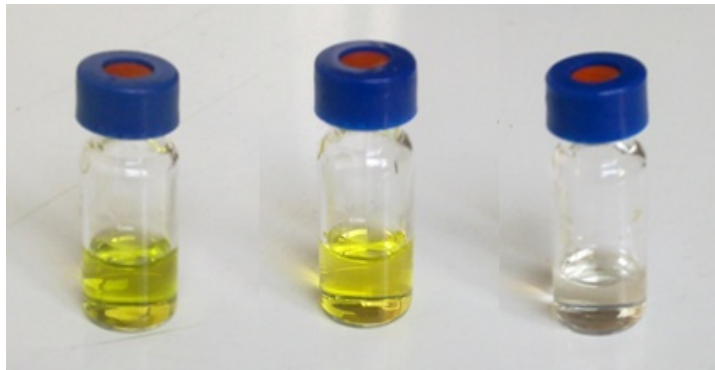


MWCNTs, amino-modified CNTs;
N-enriched Carbon

mixed with PSA/GCB/C18/PSA etc

I. d-SPE method optimization

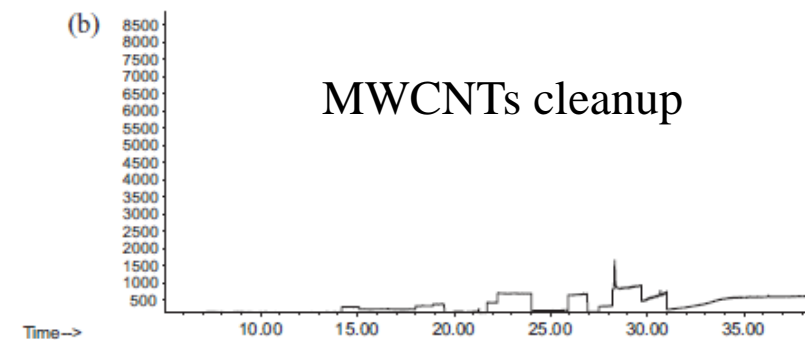
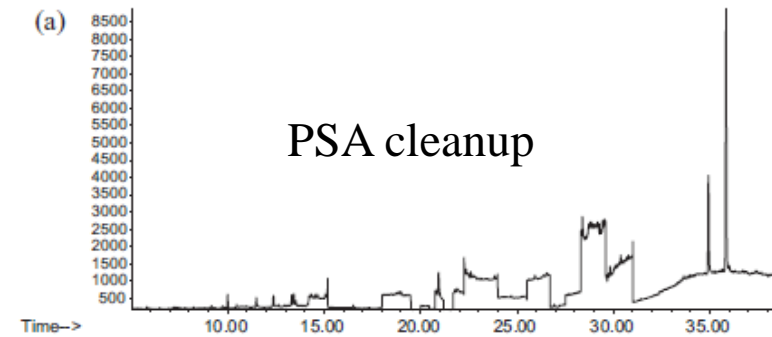
- MWCNTs as r-DSPE cleanup material for cabbage, spinach, orange and grape



**Without
cleanup**

**r-dSPE
(PSA)**

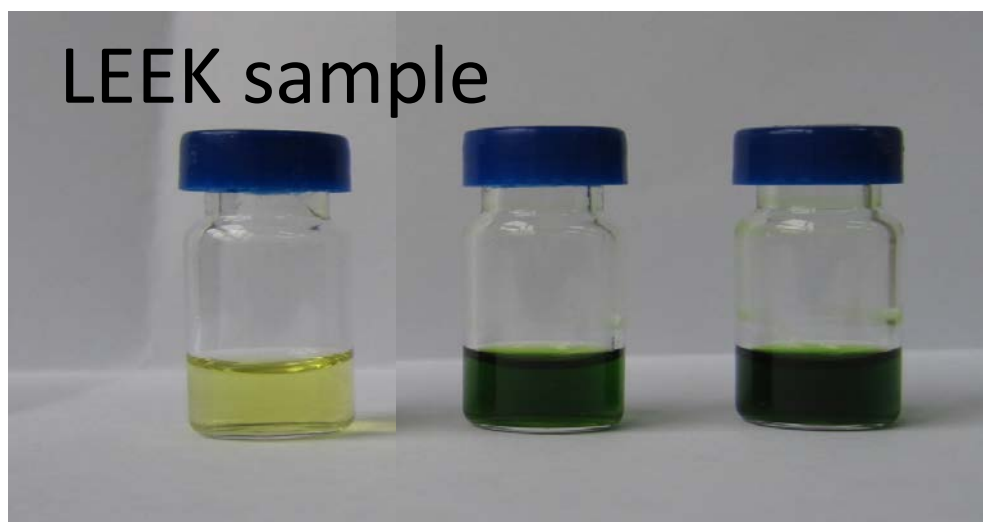
**r-dSPE
(MWCNTs)**



**MWCNTs, better cleanup performance
fewer interference peaks in the chromatogram
Method performances meet the criteria**

I. d-SPE method optimization

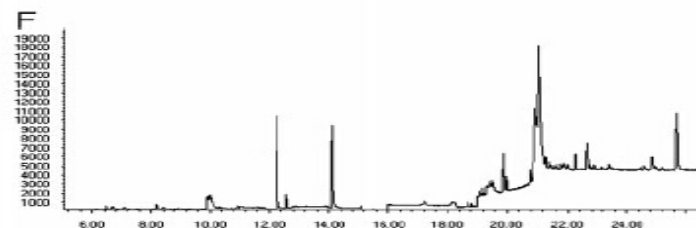
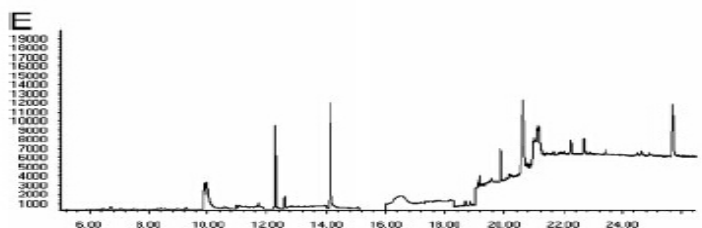
- MWCNTs as r-DSPE cleanup material for complex matrices: **leek, onion, ginger and garlic**



A: MWCNTS E:PSA F:without



Agilent GC 6890 with mass spectrometric detector



Chromatogram for blank leek extract after different r-DSPE sorbents with GCMS:

- (A) MWCNTs;
- (E) PSA;
- (F) without r-DSPE

II. m-PFC method development

Multi-residue analysis –GCMS in Soybeans by DPX method

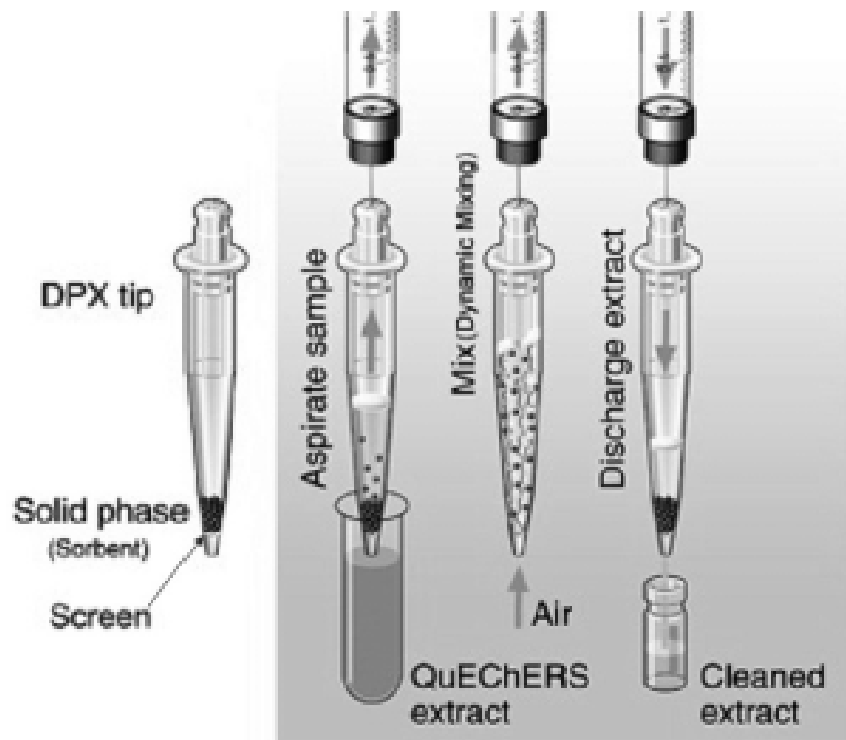


Figure 2. Schematic diagram of a DPX cleanup. Modified schematic diagram adapted from the online publication of GERSTEL, <http://www.gerstel.de/pdf/p-gc-an-2009-01.pdf> (Guan, H.; Brewer, W. E.; Morgan, S. L.; Stuff, J. R.; Whitecavage, J. A.; Foster, F. D. Automated Multi-Residue Pesticide Analysis in Fruits and Vegetables by Disposable Pipette Extraction (DPX) and Gas Chromatography/Mass Spectrometry. 2009, AN/2009, 1–7).

Ziang Li, PAN C et al, J Agric Food Chem. **2012**, 16;60(19):4788-98.

Multiresidue Analysis of 58 Pesticides in Bean Products by Disposable Pipet Extraction (DPX) Cleanup and Gas Chromatography–Mass Spectrometry Determination

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⁴College of Chemistry and Molecular Engineering, Peking University, Beijing, China

Supporting Information

ABSTRACT: A method based on disposable pipet extraction (DPX) sample spectrometric detection by selected ion monitoring (GC/MS-SIM) was established for the multiresidue analysis of 58 pesticides in soybean, mung bean, adzuki bean, and black bean. Samples were extracted with acetonitrile and aspirated into DPX tubes. Cleanup procedure was achieved in a simple 1-min process. The limits of quantification (LOQ) of this method ranged from 0.02 to 0.1 mg kg⁻¹ on soybean, mung bean, adzuki bean, and black bean were 70.2–109.1%, respectively, for all studied pesticides. Moreover, pesticide risk assessment for a Beijing market area was conducted. A maximum 0.958% of ADI (acceptable daily intake) and 55.1% of ARD (acute reference dose) for NESTI (national estimate of these products).

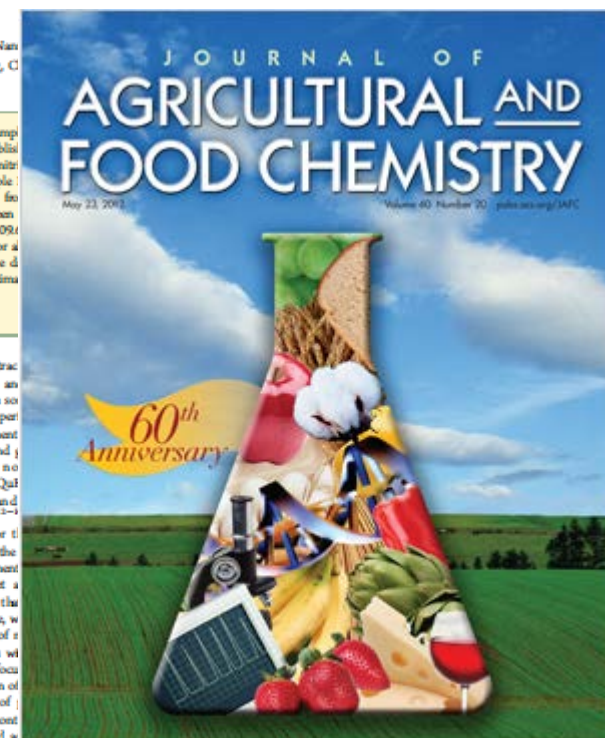
KEYWORDS: multiresidue, DPX, GC/MS, beans, pesticide, analysis

INTRODUCTION

Bean products are one of the fundamental foods recommended by the health food pyramid. Acting as both grains and vegetables in prevention of cancer, they are rich in protein and vitamins but low in calories.¹ However, public concern over food safety, especially the potential health risk to humans caused by the high levels of pesticide residues, has increased. Measuring the trace levels of pesticide residues in beans by a simple, reliable, and environmentally friendly method is becoming important. It is a particularly challenging task to test routinely and comprehensively the multiresidue pesticides in large amounts of sample matrix components that may cause false positive results. In order to clean these components up, conventional liquid–liquid extraction (LLE) is time-consuming and laborious and usually involves significant glassware usage and disposal of large volumes of hazardous organic waste.² Therefore, solid phase extraction (SPE) techniques are worth considering because of their selectivity, capability to preconcentrate pesticide, high efficiency of using organic solvents and a variety of the adsorbent materials in the SPE column.^{3,4} Furthermore, prior to chromatographic analysis, SPE cartridges have extended the application of SPE techniques for extracting and concentrating pesticides in a broad range of sample matrices.^{5–7} However, SPE often requires separate optimization for different analyte types and may not extract different classes of pesticides in foods with the same efficiency.⁸ Besides LLE and SPE, there are some new approaches, such as solid phase

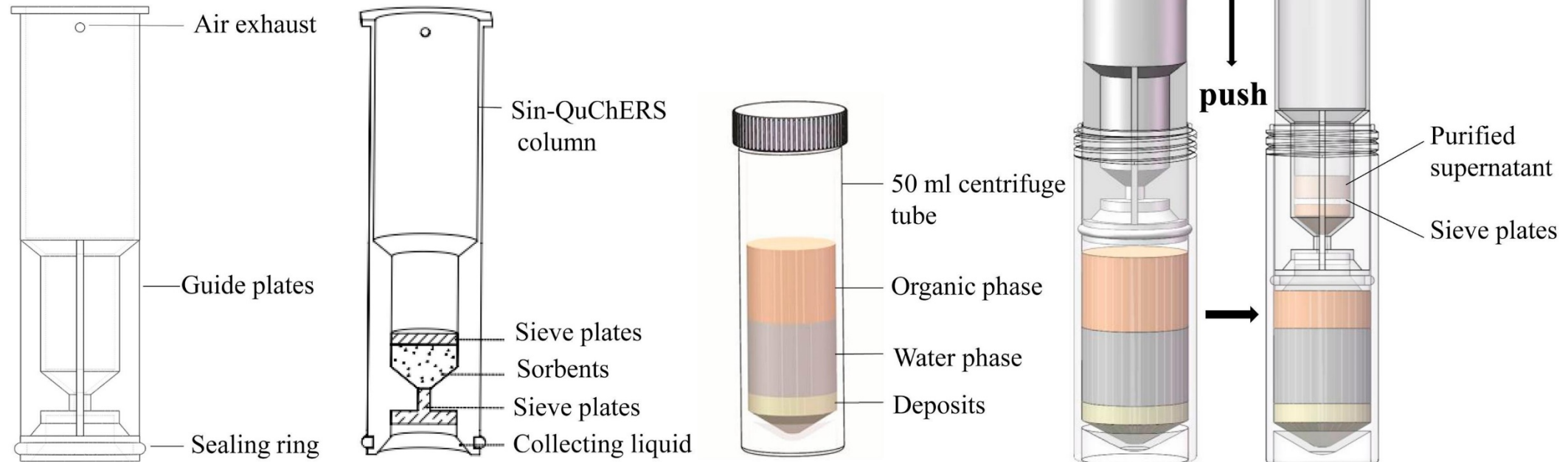
microextraction (MSPE) and matrix solid phase dispersion (MSPD) and so on, to match so their important requirements of simplicity, reliability, and low cost. The QuEChERS method is a simple, reliable, and environmentally friendly method for multiresidue analysis of pesticides in large amounts of sample matrix components that may cause false positive results. In order to clean these components up, conventional liquid–liquid extraction (LLE) is time-consuming and laborious and usually involves significant glassware usage and disposal of large volumes of hazardous organic waste.² Therefore, solid phase extraction (SPE) techniques are worth considering because of their selectivity, capability to preconcentrate pesticide, high efficiency of using organic solvents and a variety of the adsorbent materials in the SPE column.^{3,4} Furthermore, prior to chromatographic analysis, SPE cartridges have extended the application of SPE techniques for extracting and concentrating pesticides in a broad range of sample matrices.^{5–7} However, SPE often requires separate optimization for different analyte types and may not extract different classes of pesticides in foods with the same efficiency.⁸ Besides LLE and SPE, there are some new approaches, such as solid phase

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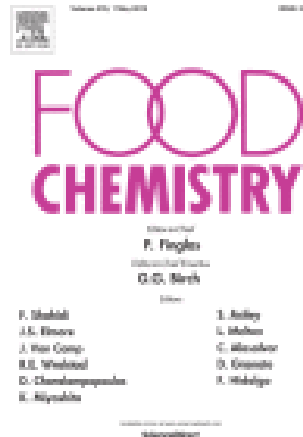


Sin-OuEChERS

Sin-QuEChERS diagram



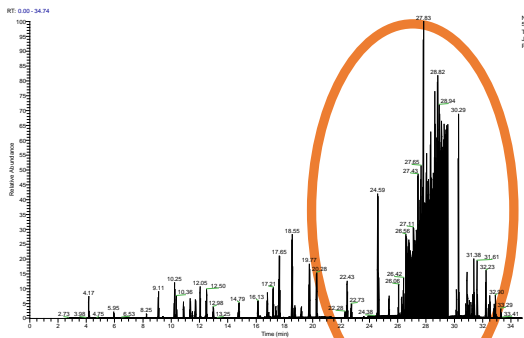
Absorbents: MWCNTS/PSA etc



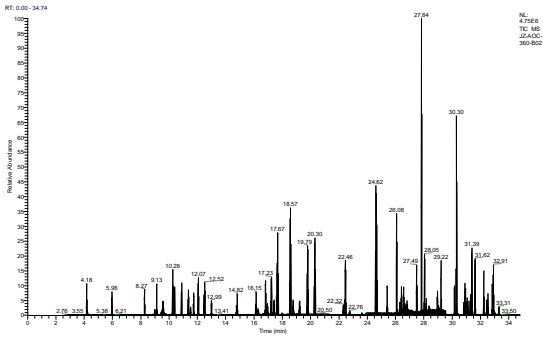
Pesticides residues in Peppers and processed products

<https://doi.org/10.1016/j.foodchem.2018.12.017>

Method specificity check comparison

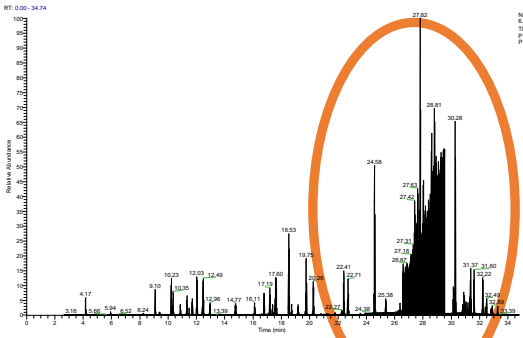


a

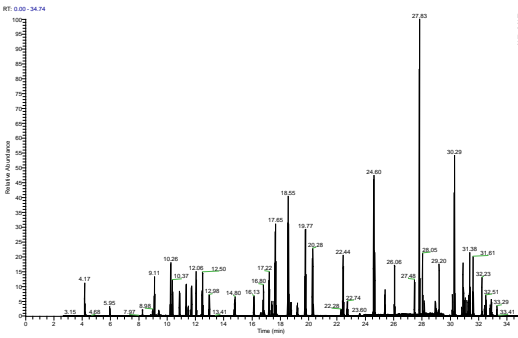


b

Citrus Sample



a



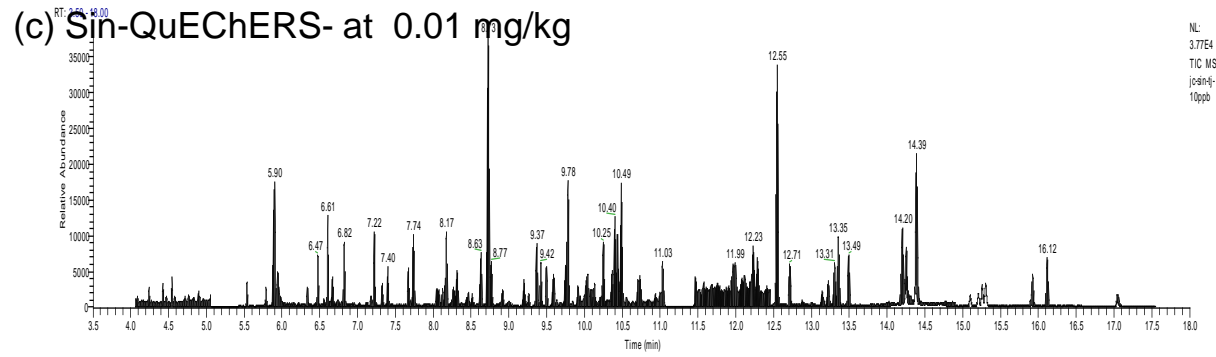
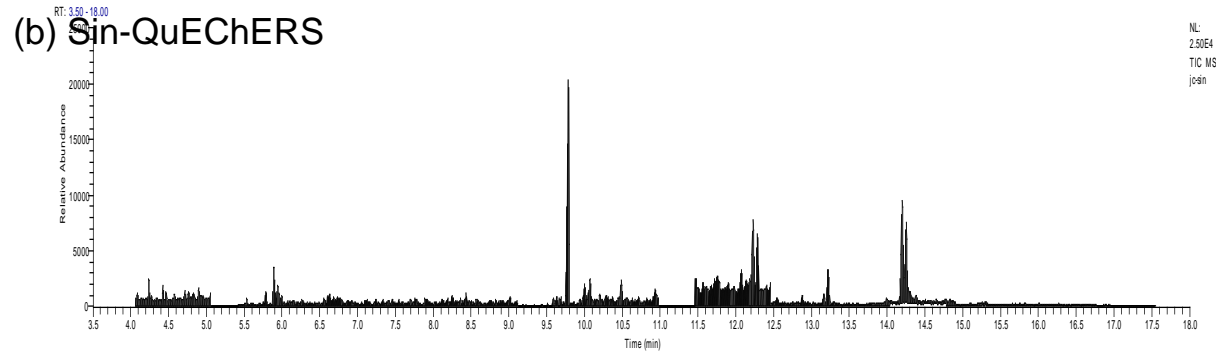
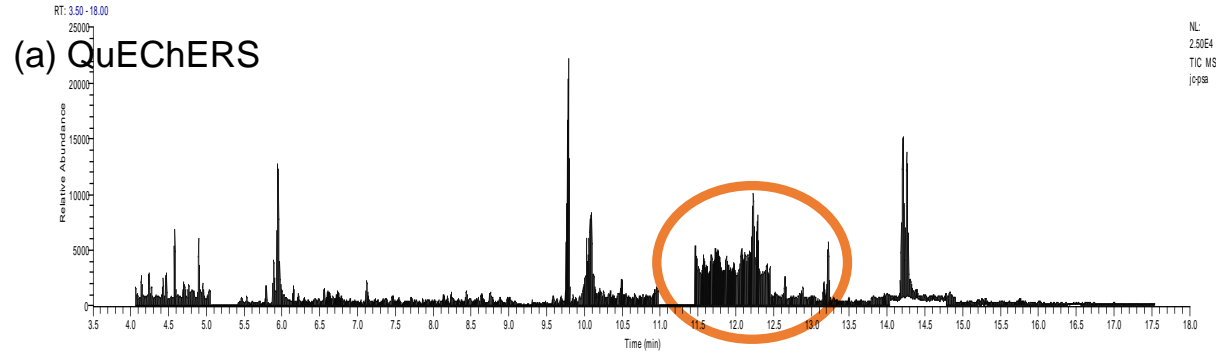
b

Grape Sample

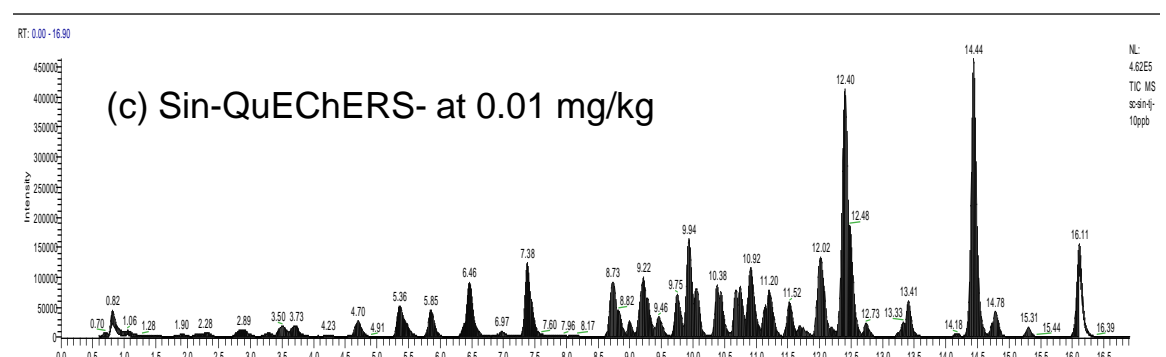
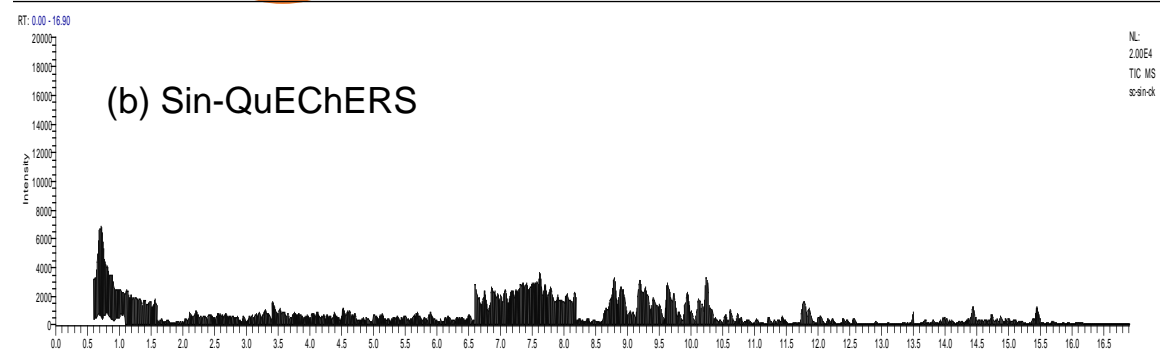
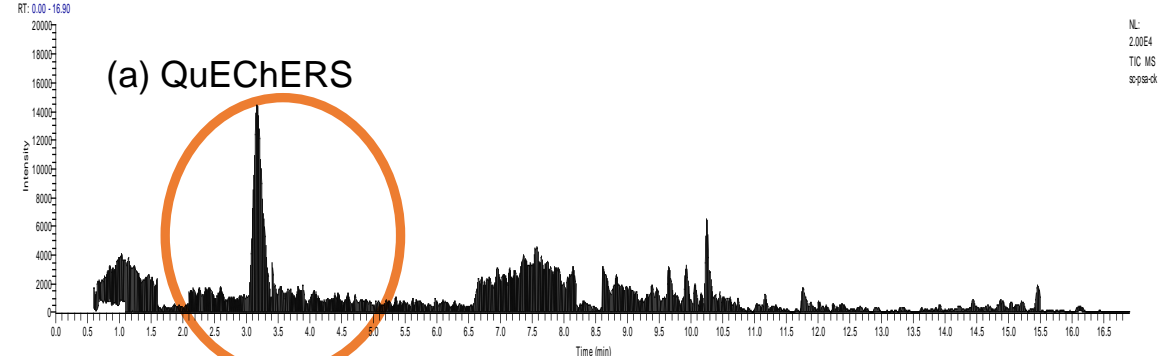
a QuEChERS

b Sin-QuEChERS-Nano

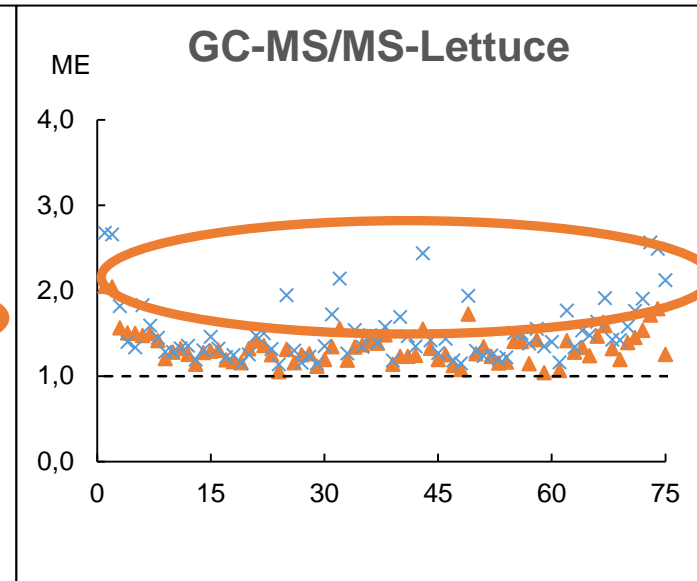
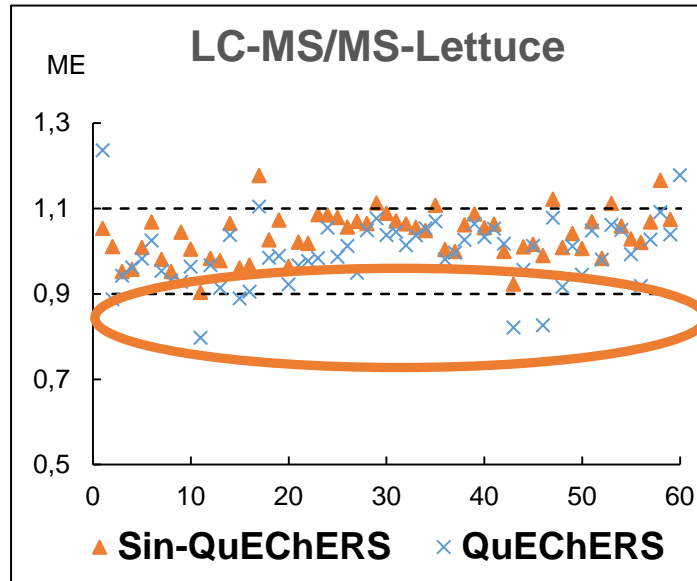
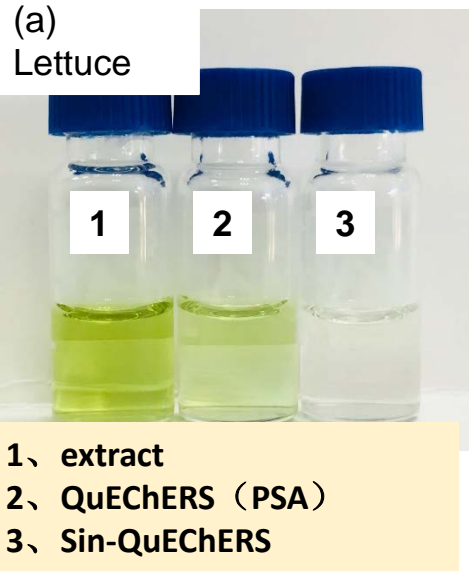
GC-MS/MS Leek



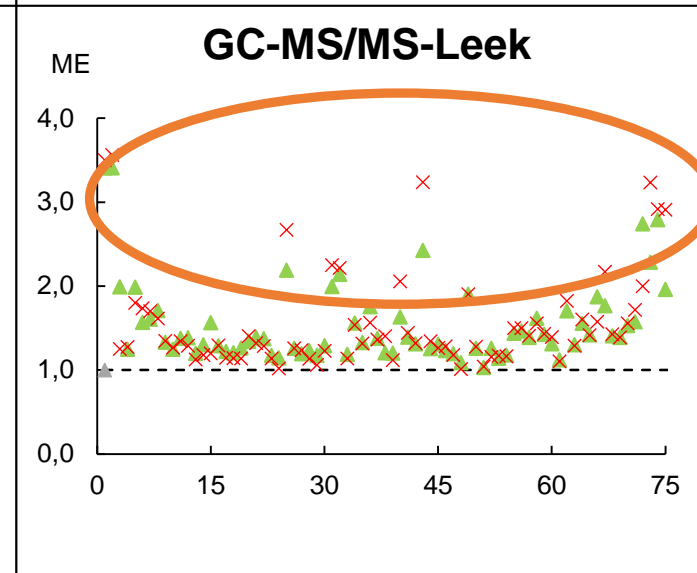
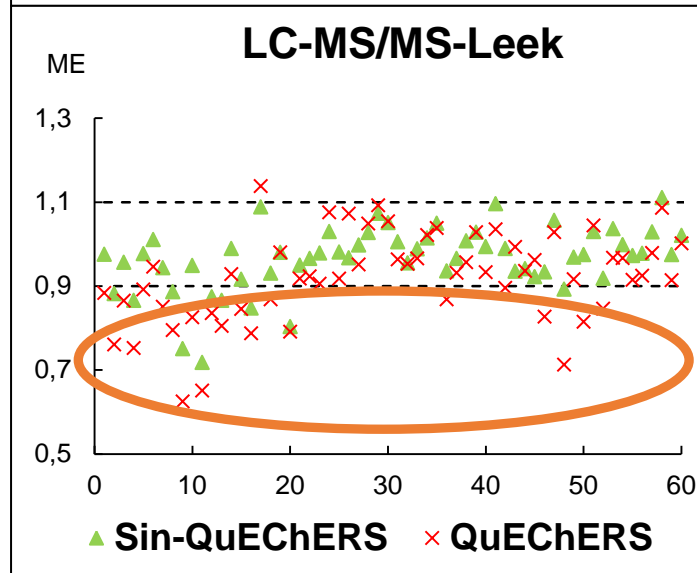
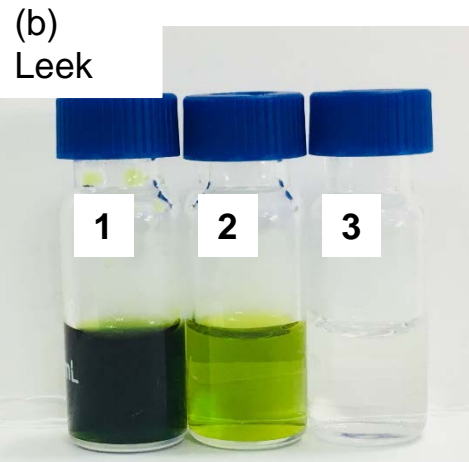
LC-MS/MS Lettuce



Matrices Effects comparison on Sin-QuEChERS –conventional QuEChERS



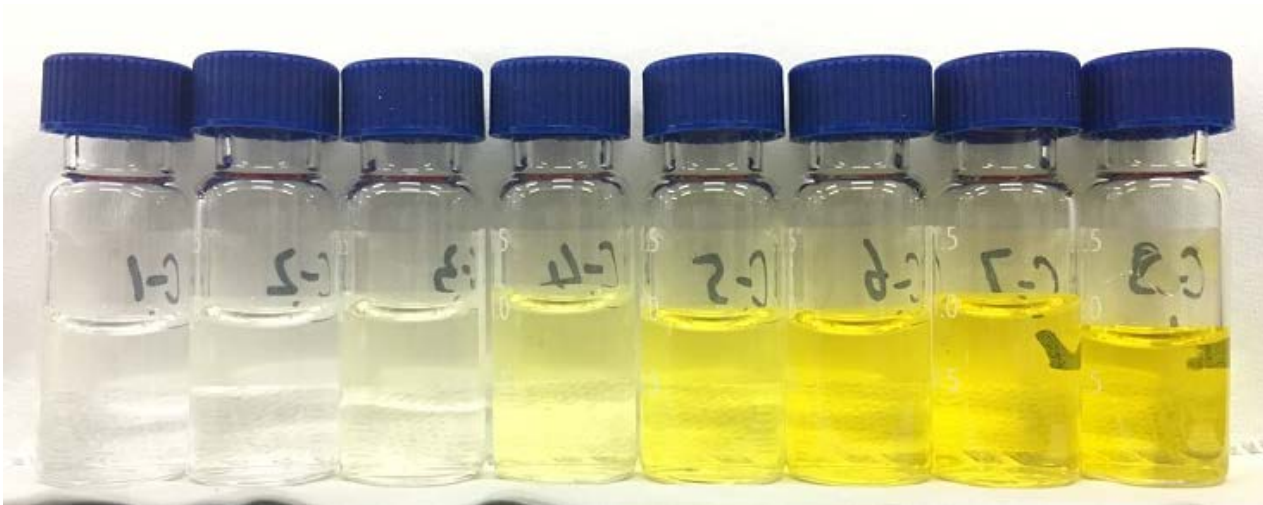
**GC-MS/MS:
75 pesticides**



**LC-MS/MS:
60 pesticides**

88 pesticides in Hemp Plant by Sin-Q; lcmsms

Sin-QuEChERS: 15mg MWCNTs+90mg PSA+80mg C18+80mg GCB+0.6g MgSO₄ + 2g NaSO₄



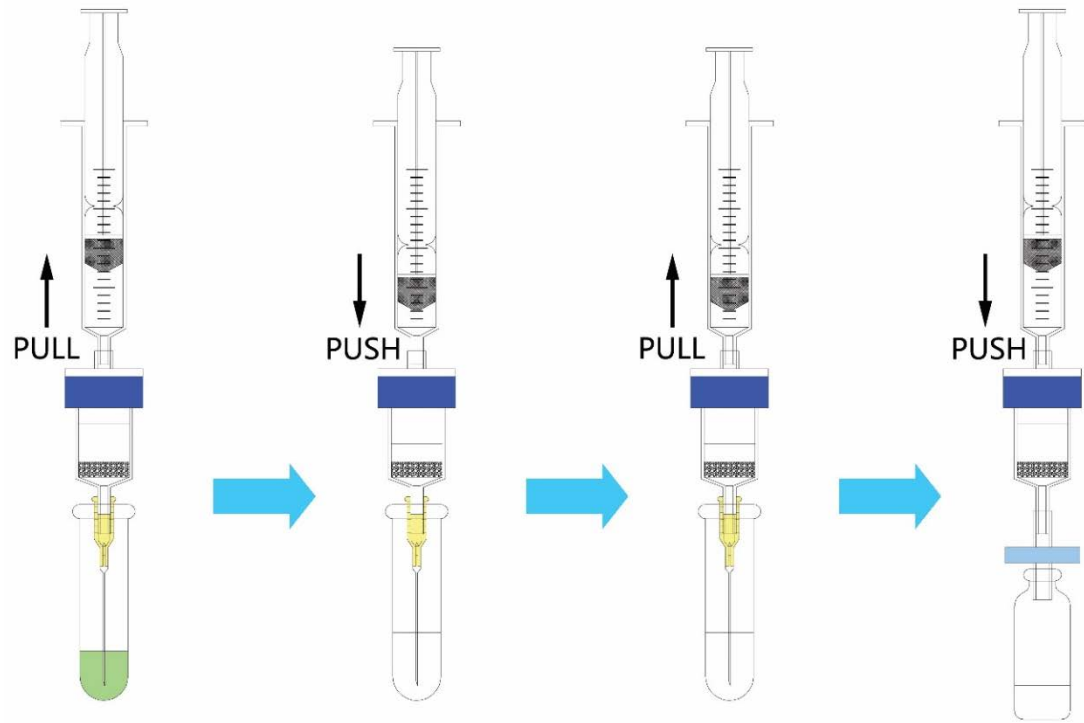
1 2 3 4 5 6 7 8

Recovery on each mL elution

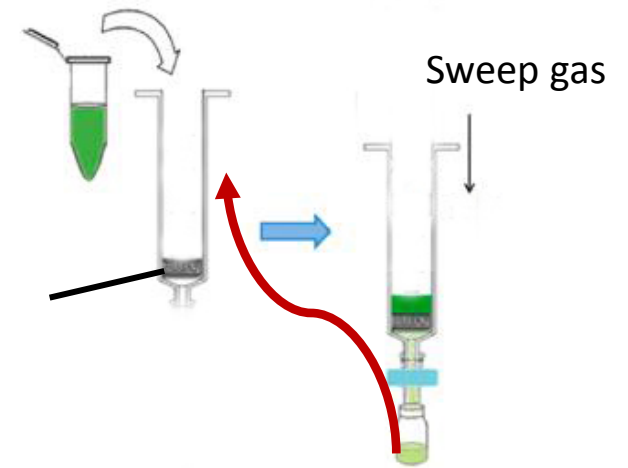
| | <70% | 70%-140% | >140% |
|------|------|----------|-------|
| 1 mL | 51 | 37 | 0 |
| 2 mL | 28 | 45 | 15 |
| 3 mL | 17 | 21 | 50 |
| 4 mL | 2 | 28 | 58 |
| 5 mL | 1 | 30 | 57 |
| 6 mL | 1 | 37 | 50 |
| 7 mL | 1 | 31 | 56 |
| 8 mL | 3 | 33 | 52 |

II. m-PFC method development

Concept of multi-competition equilibration of Matrix VS Analytes



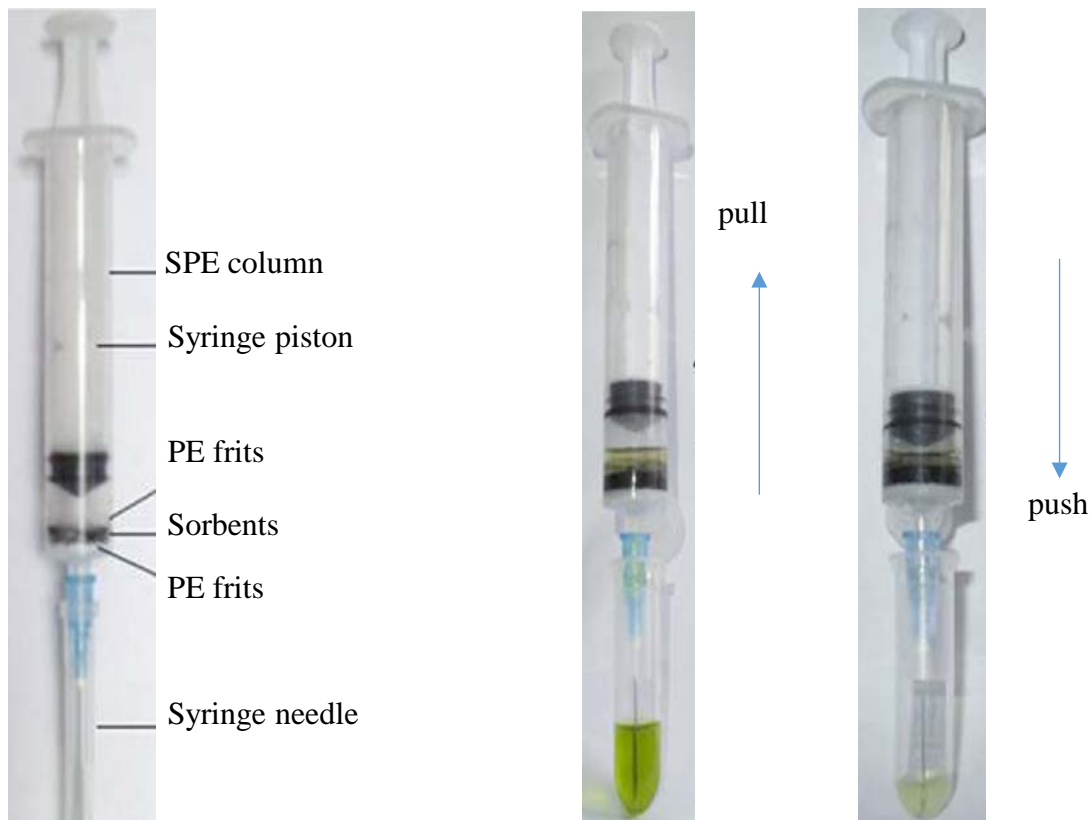
m-PFC: Multi-plug Filtration Clean-up



Multi-Filtration Cleanup : m-FC

II. m-PFC method development

Multi-plug filtration cleanup (m-PFC) for analysis of pesticide residues.



M-PFC procedure

Packed optimized d-SPE sorbents in columns, connect to syringe. Pull and push the piston to let extracts through the sorbents.

Advantages:

without solvent evaporation, vortex, centrifugation

Absorbent active sites;

multiple cycles : multiple equilibration

Improve method performance and efficacy

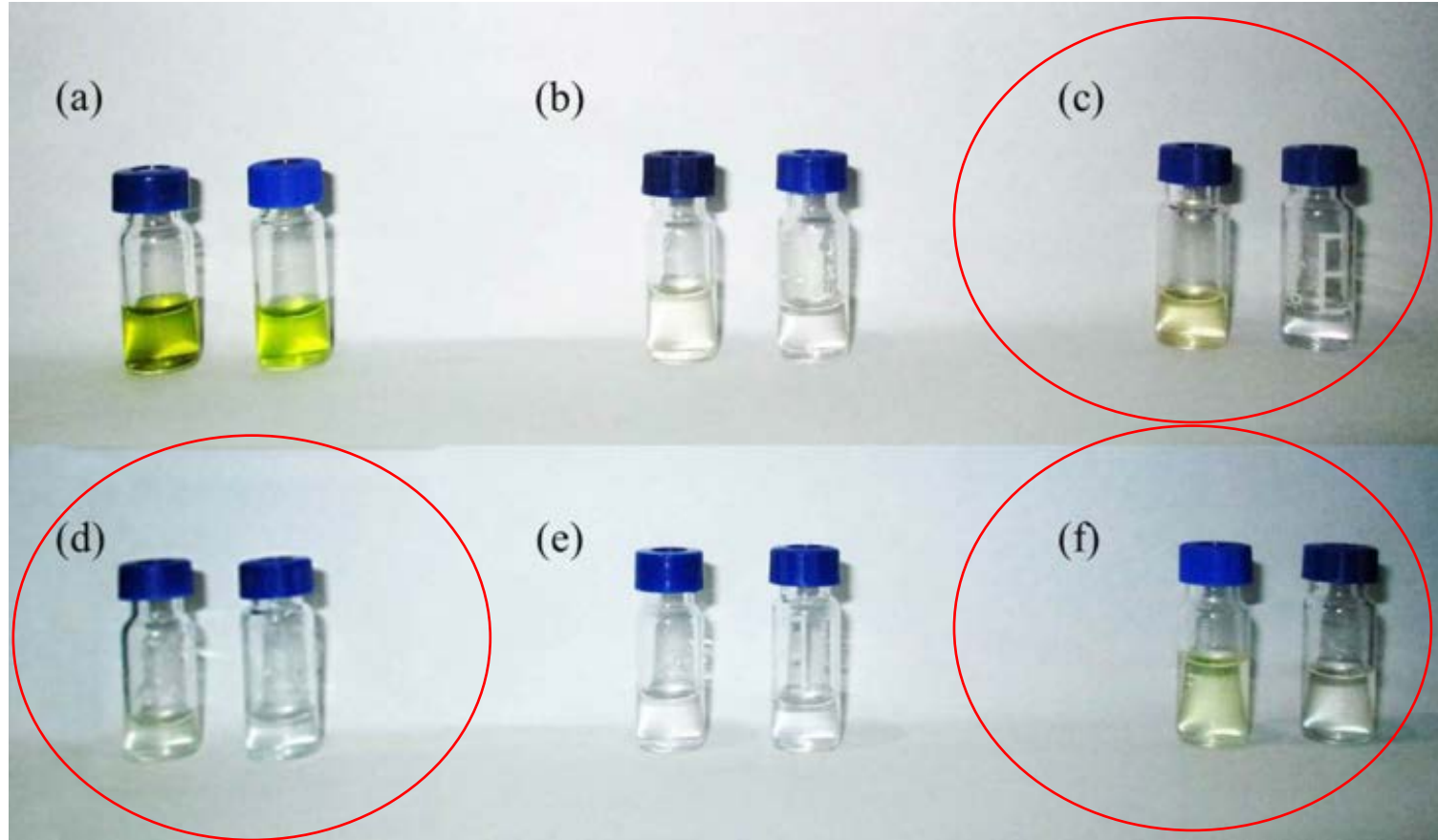
m-PFC method developemnt

Comparison: m-PFC and d-SPE were compared for:

- 25 typical **pesticides**
- in **6 matrices** (wheat, spinach, carrot, apple, citrus and peanut)
- by LC-MS/MS detection

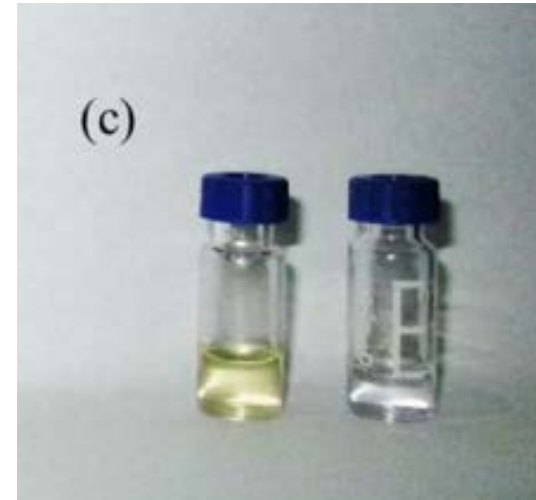
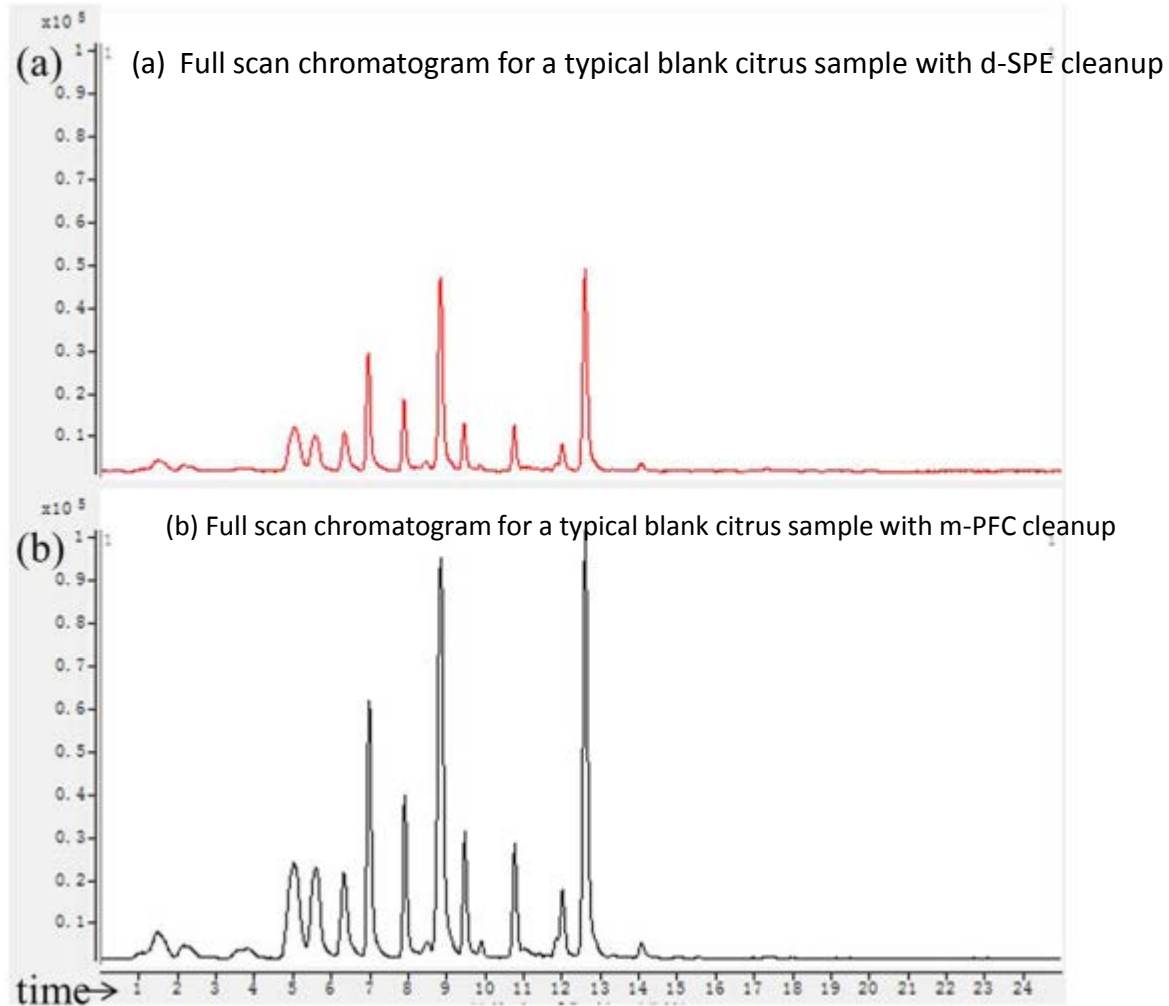


Comparison between d-SPE and m-PFC: same amount of absorbents



Comparison of visual check (left : d-SPE, right :m-PFC):
(a): spinach sample; (b): wheat sample; **(c): citrus sample;**
(d): peanut sample; (e): apple sample; **(f): carrot sample.**

Comparison between d-SPE and m-PFC

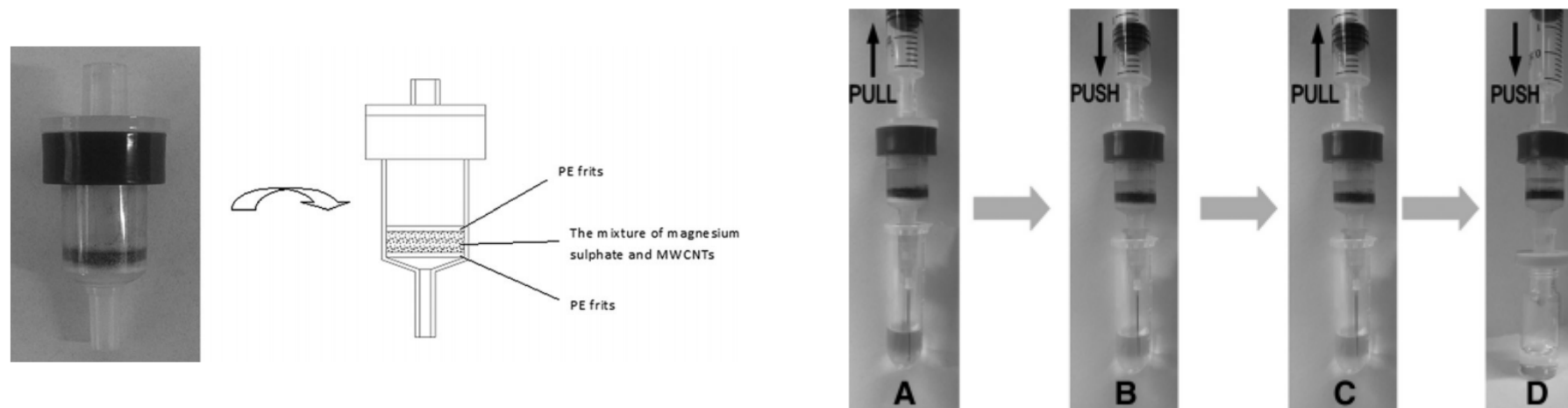


the chromatographic response intensity (Y-axis) of m-PFC method could be 2 times as of less matrix interferences

Better cleanup performance

II. m-PFC method application

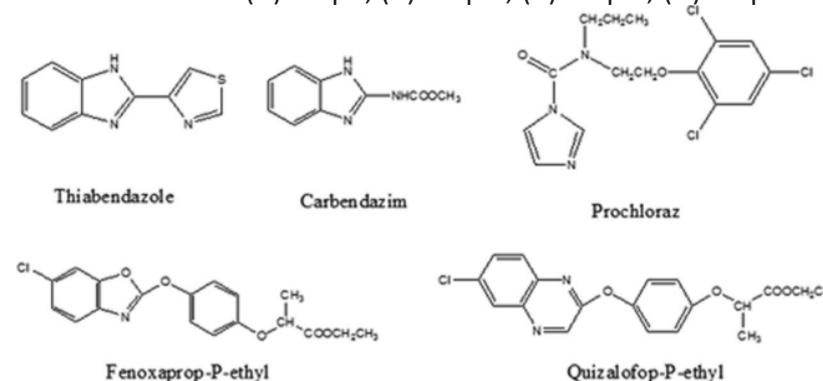
Analysis of 40 typical pesticide residues in **apple, cabbage, and potato**



Schematic diagram of the rapid m-PFC method. (A) Step1; (B) Step 2; (C) Step 3; (D) Step 4.

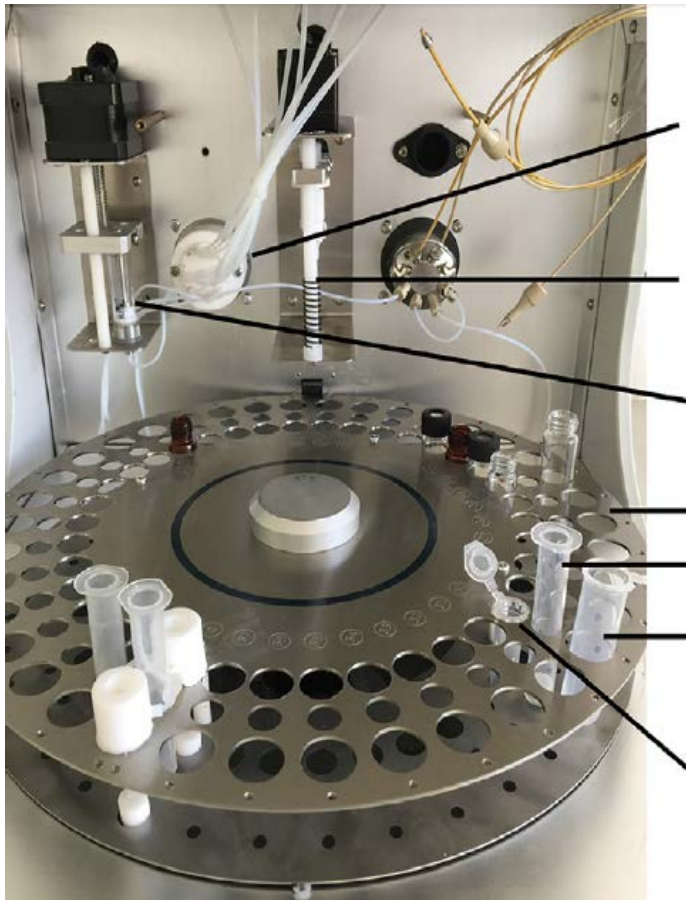
Recoveries : 71 %- 117 % (except the benzo-heterocyclic compounds)

RSDs: < 15 %



m-FC automatic device and evaluation

——Semi-automatic m-FC



Selection valve

A series of injection components

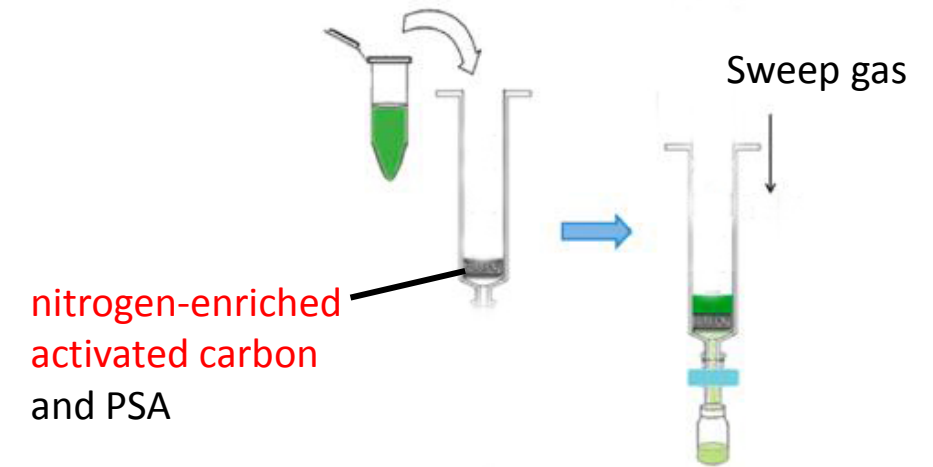
Pump syringe

Tray

m-FC column

Tube for extracts before cleanup

Tube for extracts after cleanup

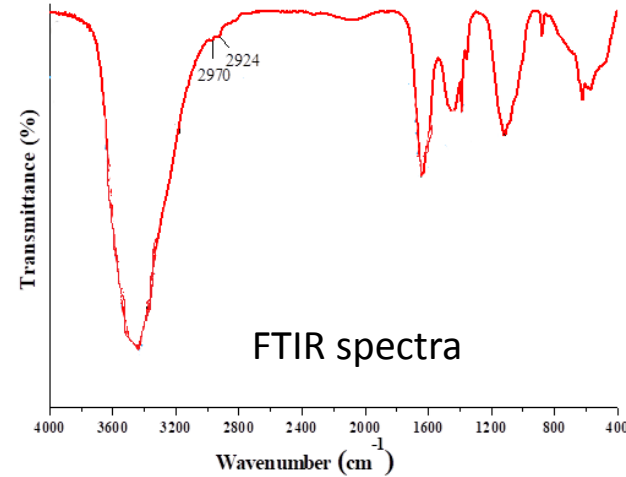
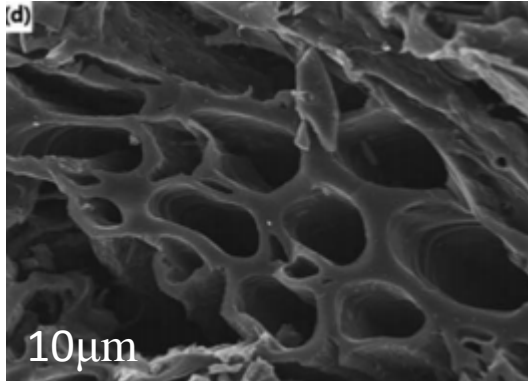


- Optimization of m-FC Cycles
- Optimization of m-FC volume
- Optimization of flow rate
- With **nitrogen-enriched activated carb**

Components diagram of the Auto m-FC equipment

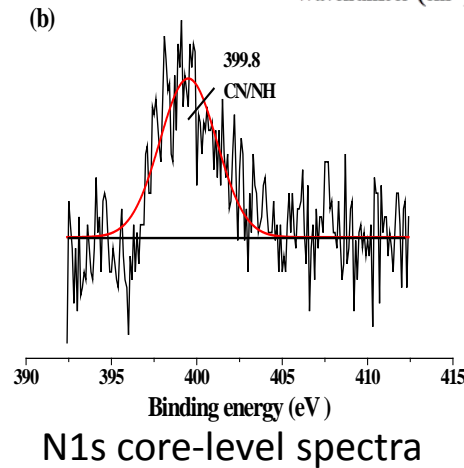
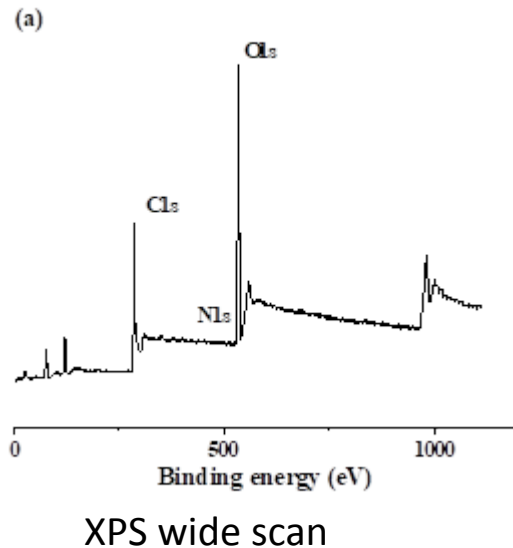
m- FC automatic device and evaluation

—Semi-automatic m-FC



nitrogen-enriched activated carbon

The BET surface area: 1868.36(m²/g)



Eco-friendly material prepared from the wasted medium density fiberboard

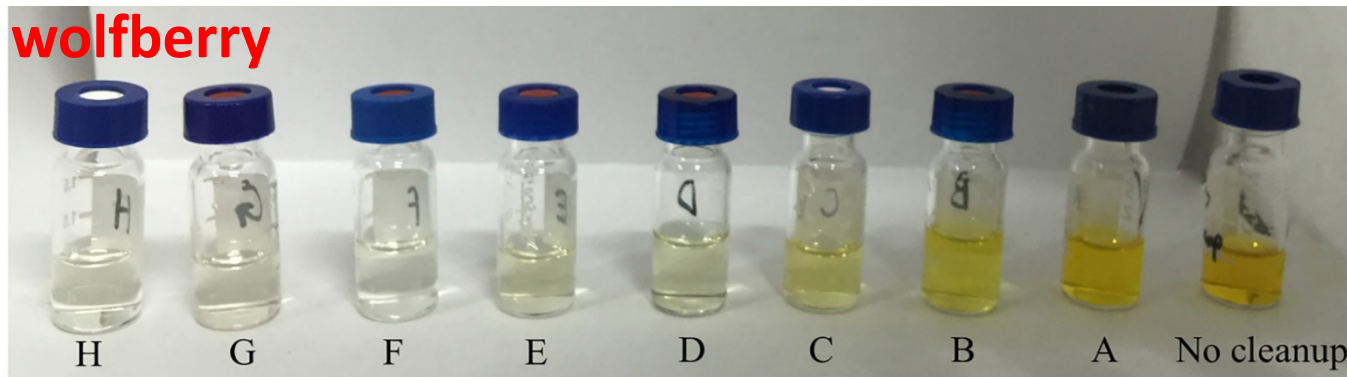
III. m- FC automatic device and evaluation

—Semi-automatic m-FC

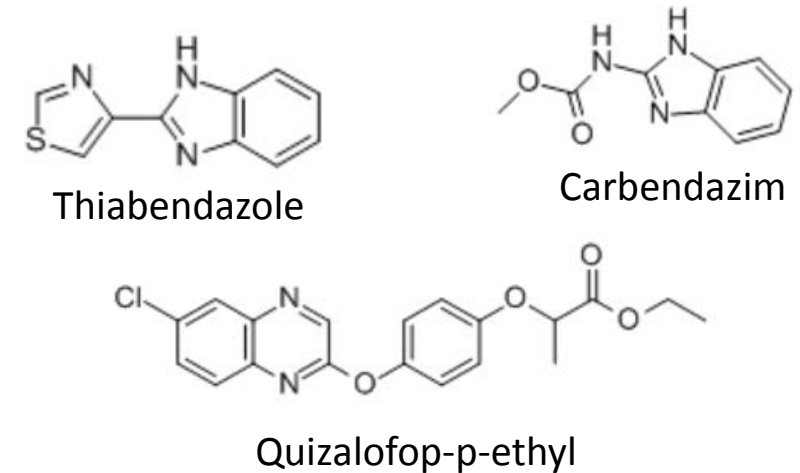
nitrogen-enriched activated carb: adsorb interfering substances in matrices, rather than the analytes

MWCNTs: some benzo-heterocyclic compounds loss

wolfberry



- A: 5 mg MWCNTs+ 30 mg PSA + 150 mg MgSO₄
- B: 5 mg MWCNTs+ 50 mg PSA + 150 mg MgSO₄
- C: 5 mg MWCNTs+ 30 mg PSA + 5 mg GCB+ 150 mg MgSO₄
- D: 5 mg MWCNTs+ 50 mg PSA + 5 mg GCB+ 150 mg MgSO₄
- E: 10 mg commercial activated carbon+ 150 mg MgSO₄
- F: 10 mg commercial activated carbon+ 50 mg PSA+ 150 mg MgSO₄
- G: 10 mg nitrogen-enriched activated carbon+ 150 mg MgSO₄
- H: 10 mg nitrogen-enriched activated carbon+ 50 mg PSA+ 150 mg MgSO₄

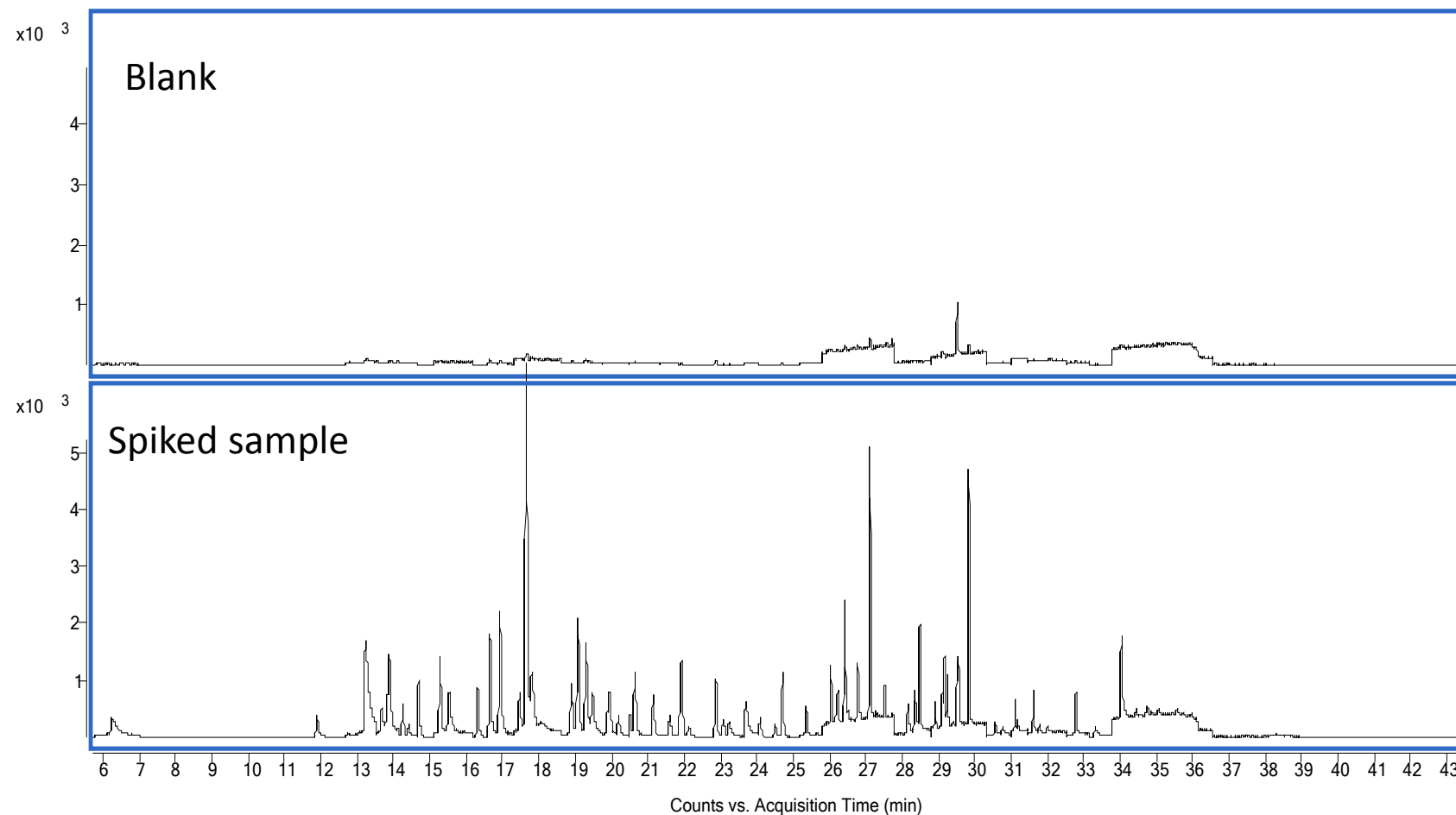
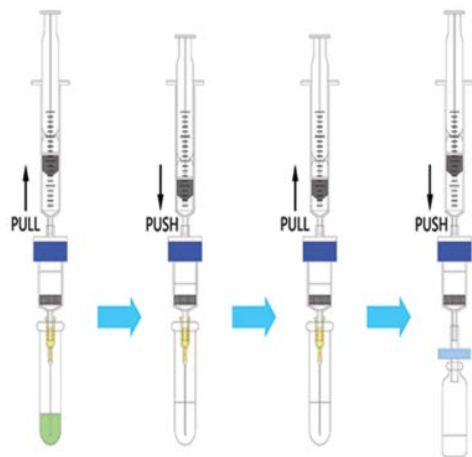


| | apple | citrus | peanut | wheat | tea | spinach |
|--------------------|--------|--------|--------|--------|-------|---------|
| Thiabendazole | 89(6) | 93(4) | 93(4) | 104(2) | 84(6) | 95(1) |
| Quizalofop-p-ethyl | 93(4) | 96(3) | 92(3) | 93(6) | 93(2) | 94(6) |
| Carbendazim | 101(3) | 89(3) | 87(6) | 89(3) | 96(7) | 88(4) |

Recoveries and RSDs at 100µg/kg in six matrices

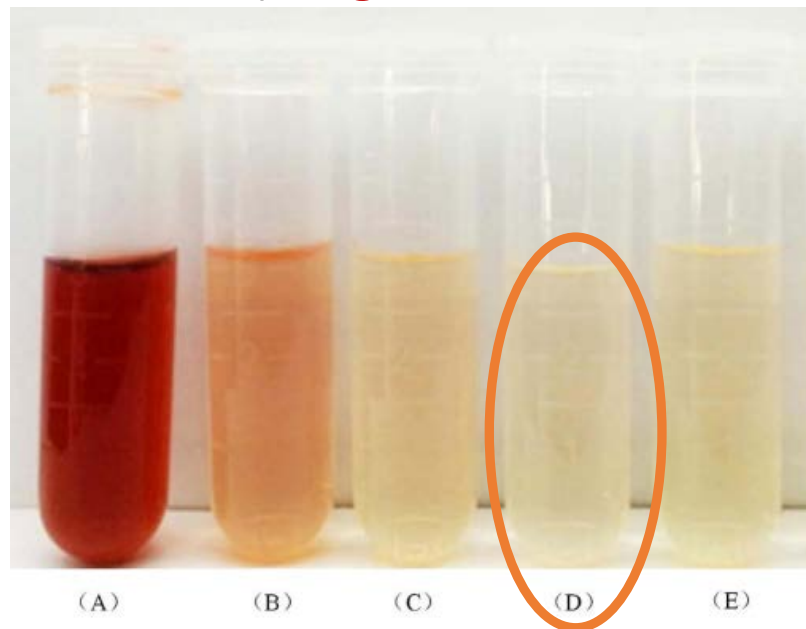
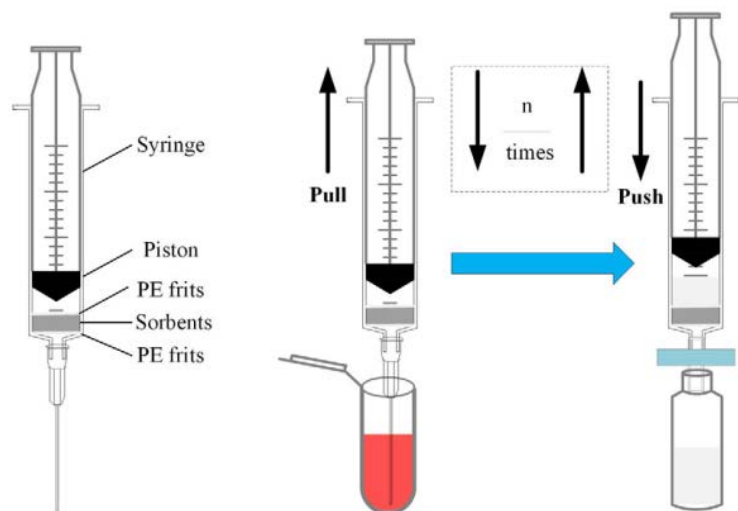
II. m-PFC method application

- Analysis of 186 pesticide residues in tomato and tomato products



II. m-PFC method application

- Analysis of 142 pesticide residues in Chinese liquor (Moutai etc) and liquor-making raw materials (**sorghum and rice hull**)



(A) without cleanup,
(B) 1 cycle,
(C) 2 cycles,
(D) 3 cycles,
(E) 4 cycles.

Method validation: $n=2-3$

Recoveries : 71 %- 121 %

RSDs: < 16.8 %

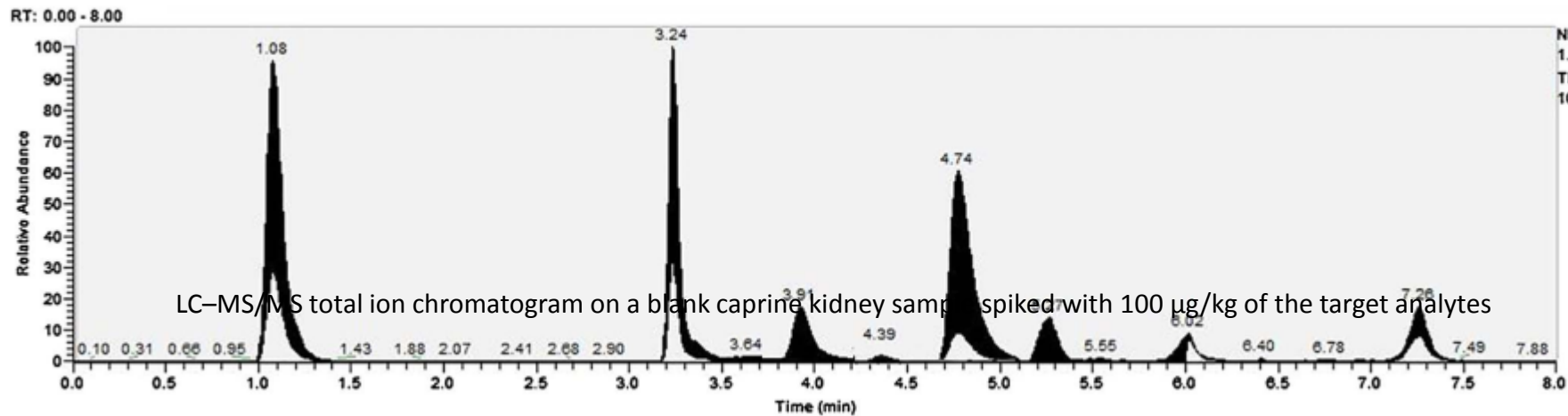
LOQs: 0.1~5 $\mu\text{g}/\text{kg}$

LODs: 0.03~1.5 $\mu\text{g}/\text{kg}$

Han Y, Song L, PAN C, et al. *Food Chemistry*, 2018:258-267.

II. m-PFC method application

- Analysis of veterinary drug residues in typical **animal matrices: meat, kidney, milk:**
- involved **low-temperature** cleanup to remove lipid, and further by **m-PFC** with MWCNTs and other



Recoveries : 82-107%

RSDs: < 15%

| Analyte | bovine meat (µg/kg) | | | caprine meat (µg/kg) | | | swine meat (µg/kg) | | | bovine kidney (µg/kg) | | caprine kidney (µg/kg) | | swine kidney (µg/kg) | | milk (µg/kg) | |
|--------------------------|---------------------|-----------------|------|----------------------|-----------------|------|--------------------|-----------------|------|-----------------------|-----------------|------------------------|-----------------|----------------------|-----------------|-----------------|-----------------|
| | CC _α | CC _β | MRL | CC _α | CC _β | MRL | CC _α | CC _β | MRL | CC _α | CC _β | CC _α | CC _β | CC _α | CC _β | CC _α | CC _β |
| <i>cis</i> -Tilmicosin | 50.1 | 50.1 | 50.0 | 50.1 | 50.3 | 50.0 | 50.2 | 50.4 | 50.0 | 1.00 | 1.26 | 1.00 | 1.18 | 1.00 | 1.25 | 0.20 | 0.36 |
| <i>trans</i> -Tilmicosin | 50.1 | 50.3 | | 50.2 | 50.5 | | 50.2 | 50.5 | | 1.00 | 1.25 | 1.00 | 1.25 | 1.00 | 1.23 | 0.20 | 0.43 |
| Sulfamethazine | 100 | 100 | 100 | 100 | 101 | 100 | 100 | 101 | 100 | 2.00 | 2.30 | 2.00 | 2.26 | 2.00 | 2.26 | 1.00 | 1.20 |
| Sulfamethoxazole | 100 | 101 | 100 | 100 | 101 | 100 | 100 | 100 | 100 | 2.00 | 2.15 | 2.00 | 2.20 | 2.00 | 2.15 | 1.00 | 1.15 |
| Sulfamonomethoxine | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 2.00 | 2.25 | 2.00 | 2.28 | 2.00 | 2.25 | 1.00 | 1.13 |
| Abamectin | 20.1 | 20.2 | 20.0 | 2.00 | 2.13 | - | 2.00 | 2.28 | - | 2.00 | 2.30 | 2.00 | 2.30 | 1.00 | 1.28 | 2.00 | 2.25 |
| Doramectin | 20.1 | 20.2 | 20.0 | 2.00 | 2.25 | - | 2.00 | 2.18 | - | 2.00 | 2.26 | 2.00 | 2.26 | 1.00 | 1.18 | 2.00 | 2.21 |
| Ivermectin | 20.1 | 20.2 | 20.0 | 2.00 | 2.23 | - | 2.00 | 2.33 | - | 2.00 | 2.33 | 2.00 | 2.15 | 1.00 | 1.20 | 2.00 | 2.20 |

CC_αs and CC_βs of the analytes in each matrix

II. m-PFC method application

- Sample preparation- with IMS and Raman rapid detection



10 g of homogenized apple sample

↓
Add 10 mL Acetonitrile

↓ Shake vigorously 1 min

Add 5 g NaCl

↓ Shake 1 min and Centrifuge

Clean up with **m-PFC**

↓
Raman Spectr.

↓
Ion Mobility Spectroscopy

→ DART-MS; ASAP-MS/MS

II. m-PFC method application

- Ion Mobility Spectrometry(IMS) for rapid analysis

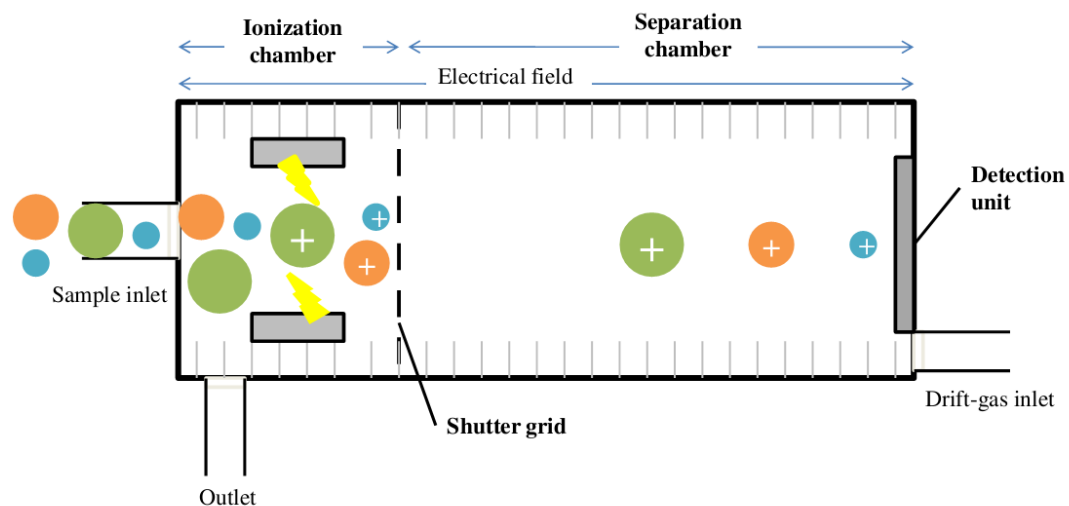
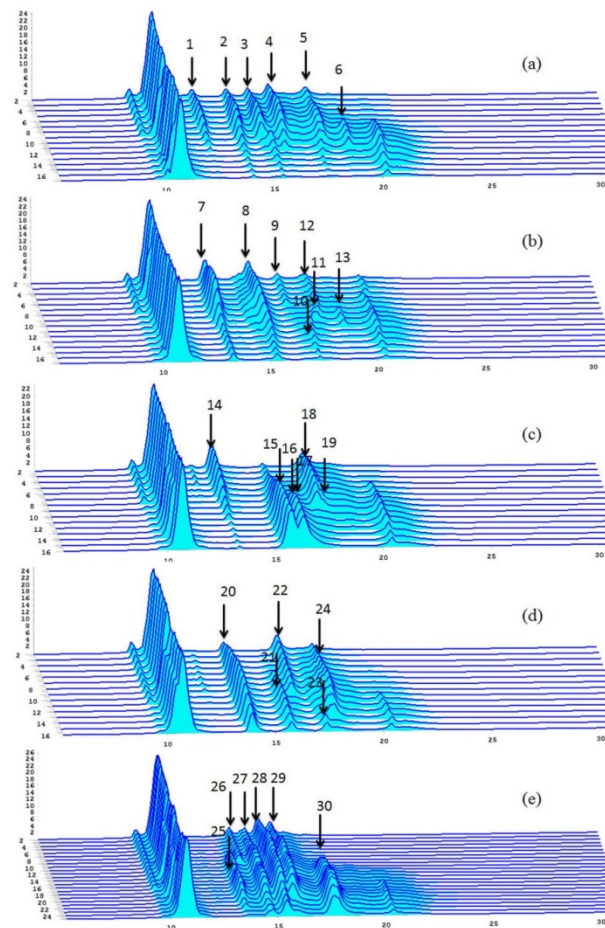
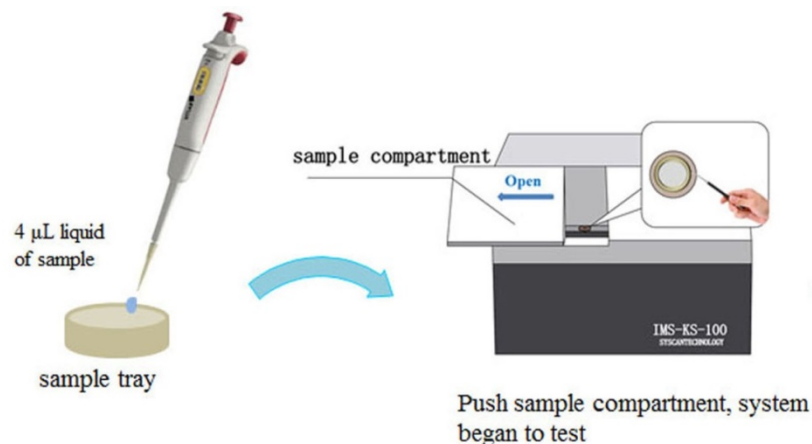
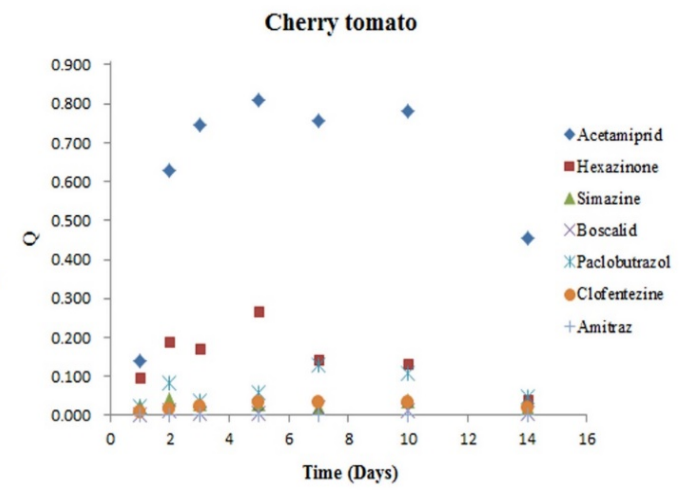
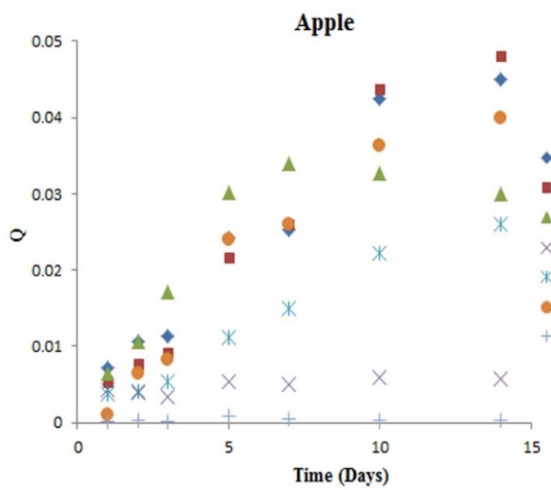
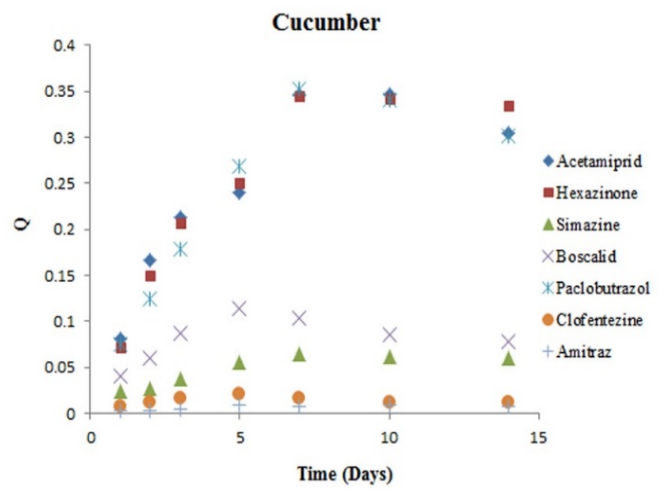
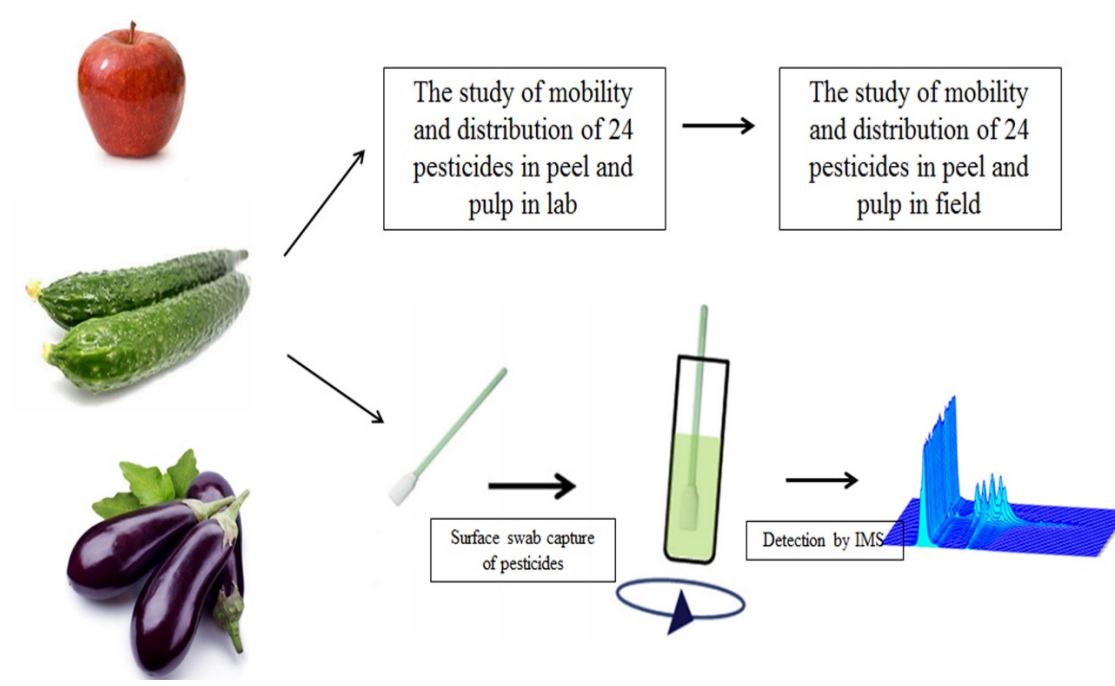
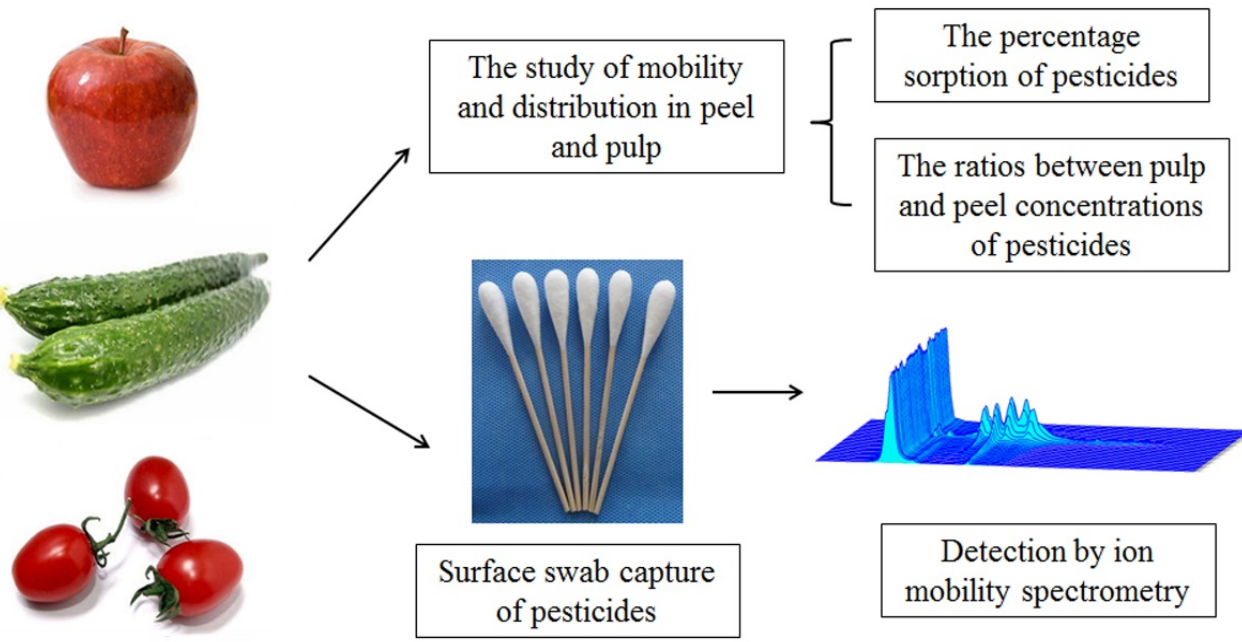


Figure 2. A conventional ion-mobility spectrometry (IMS) device.



- (a) 1 Methamidophos, 2 Omethoate, 3 Monocrotophos, 4 Isocarbophos, 5 Phosphamidon, 6 Triadimenol;
- (b) 7 Methomyl, 8 Carbaryl, 9 Isoprocarb, 10 Metolachlor, 11 Chlorbenzuron, 12 Uniconazole, 13 Triadimefon;
- (c) 14 Acephate, 15 Acetamiprid, 16 Thiacloprid, 17 Nitenpyram, 18 Paclobutrazol, 19 Propiconazol;
- (d) 20 Tricyclazole, 21 Dimethomorph, 22 Propyzamide, 23 Tebuconazole, 24 Myclobutani;
- (e) 25 Diazinon, 26 Simazine, 27 Atrazine, 28 Ametryn, 29 Alachlor, 30 RH-5849.



II. m-PFC method application

Different cleanup methods in a blank apple sample

- Raman for rapid analysis for HHPs

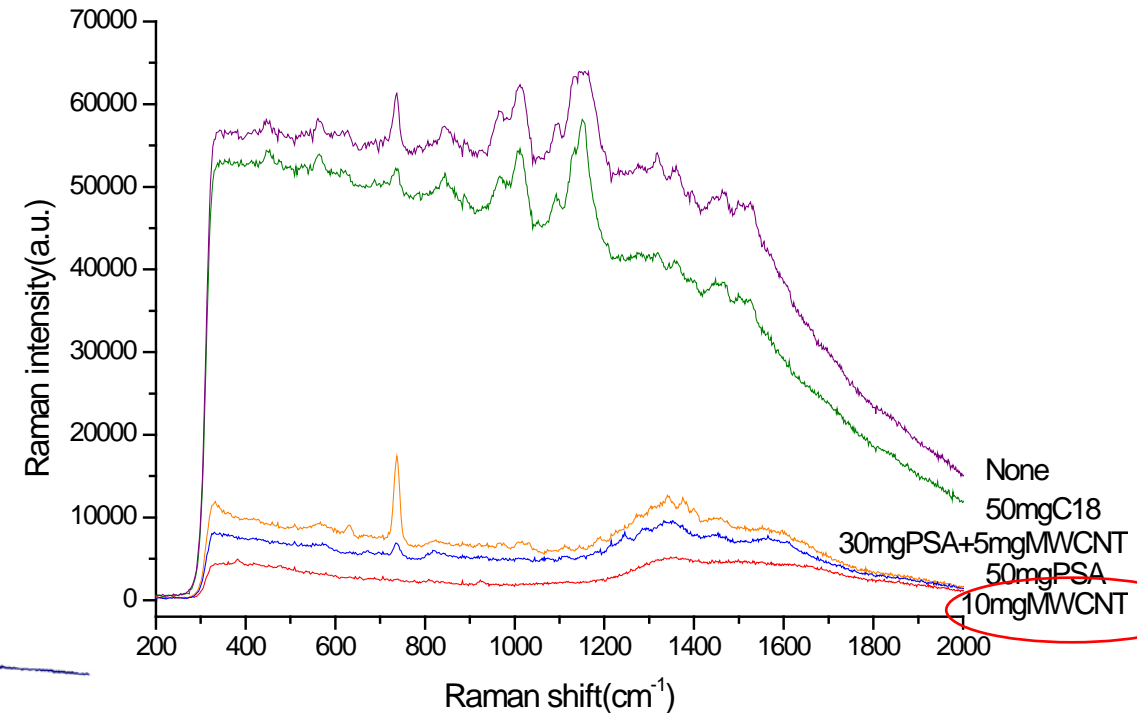
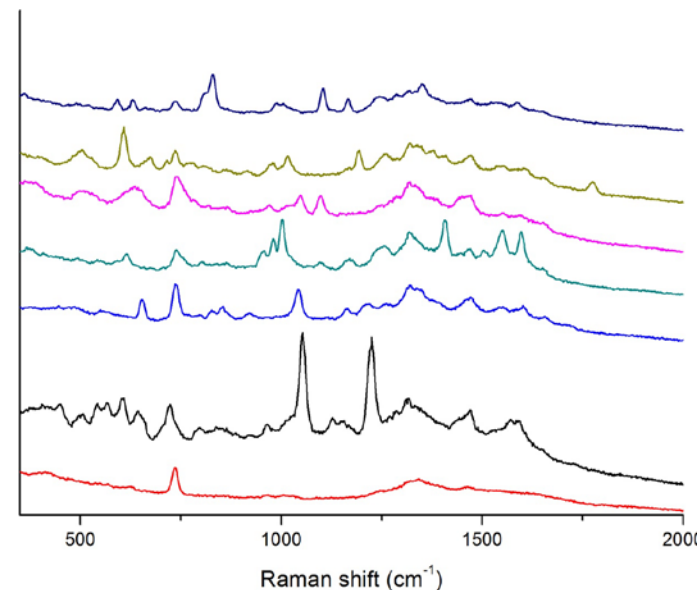


Traditional methods

- Long time
- Hard to real-time detection

Raman spectroscopy

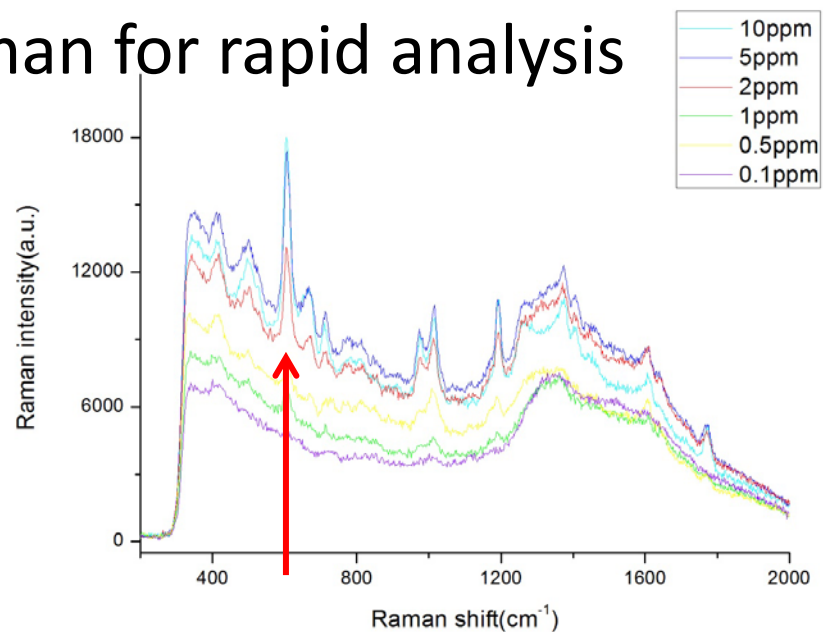
- Detect in seconds
- Small and portable



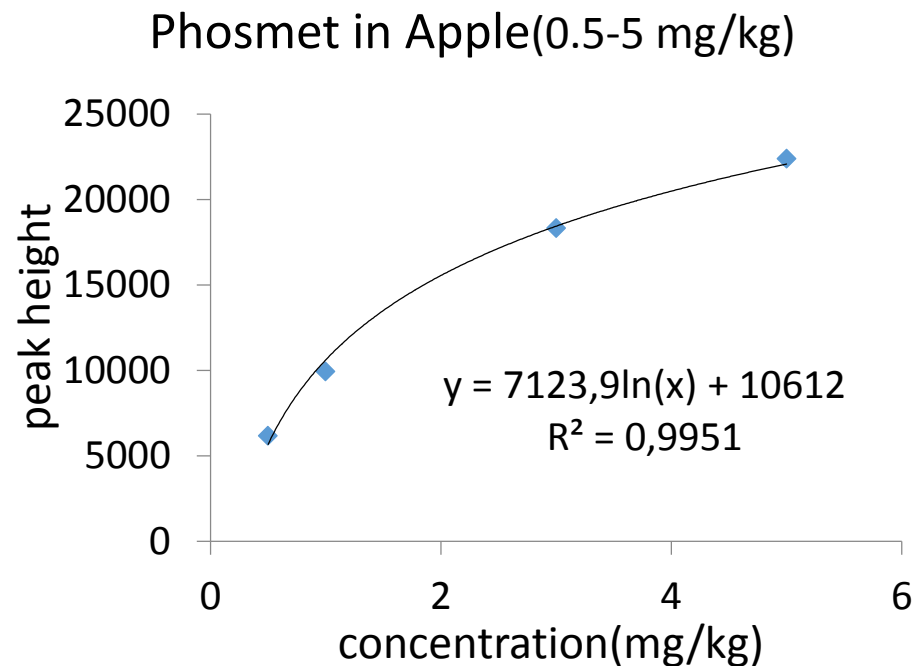
blank

II. m-PFC method application

- Raman for rapid analysis



Quantification

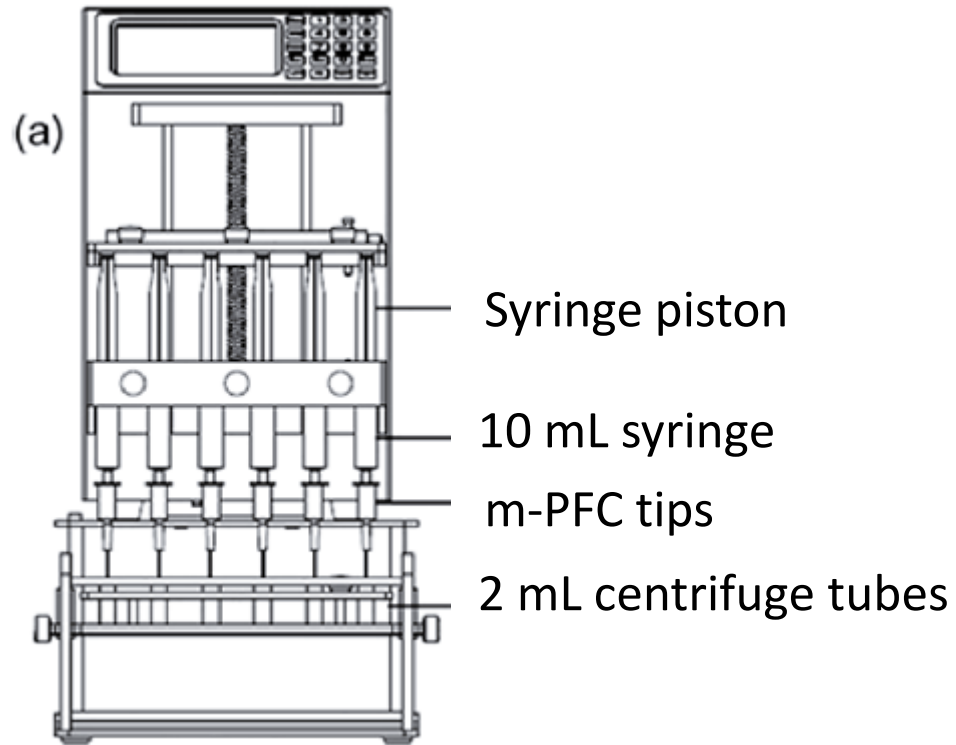


Method Recoveries

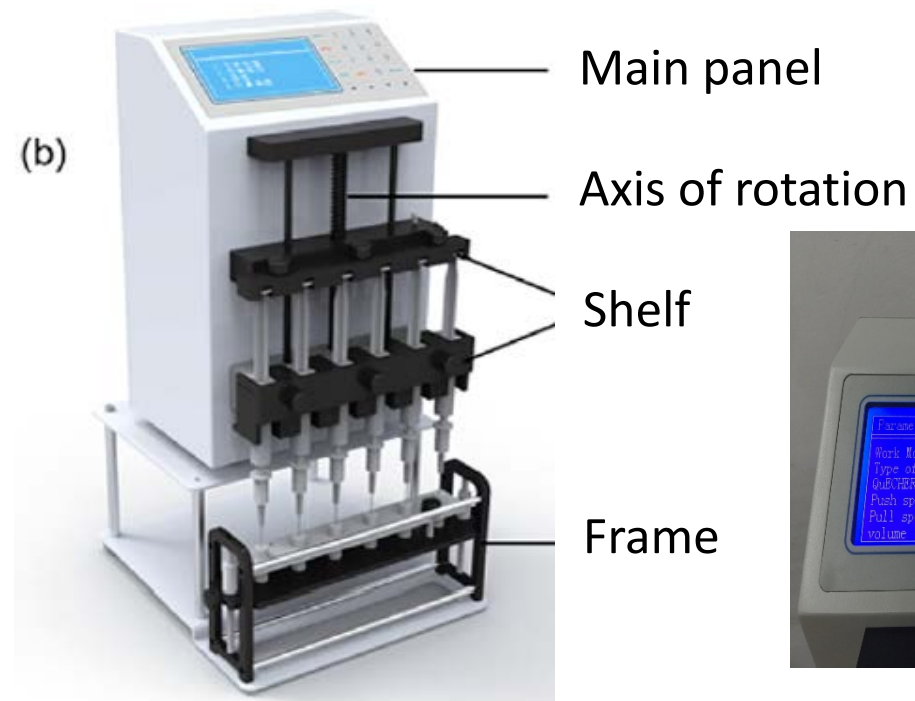
| concentration (mg/kg) | recovery % (n=4) | | | | average recovery % | RSD% |
|--------------------------|------------------|--------|--------|--------|-----------------------|------|
| | 1 | 2 | 3 | 4 | | |
| 0.5 | 83.1% | 101.4% | 101.1% | 100.4% | 96.5% | 9.3 |
| 1 | 73.3% | 73.1% | 93.3% | 93.5% | 83.3% | 14 |
| 2 | 71.8% | 84.1% | 77.3% | 75.8% | 77.3% | 6.6 |

III. m-PFC automatic device and evaluation

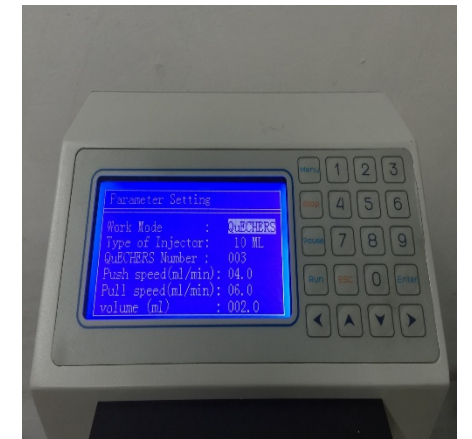
——Semi-automatic m-PFC



(a) schematic diagram of equipment



(b) components of automated m-PFC equipment

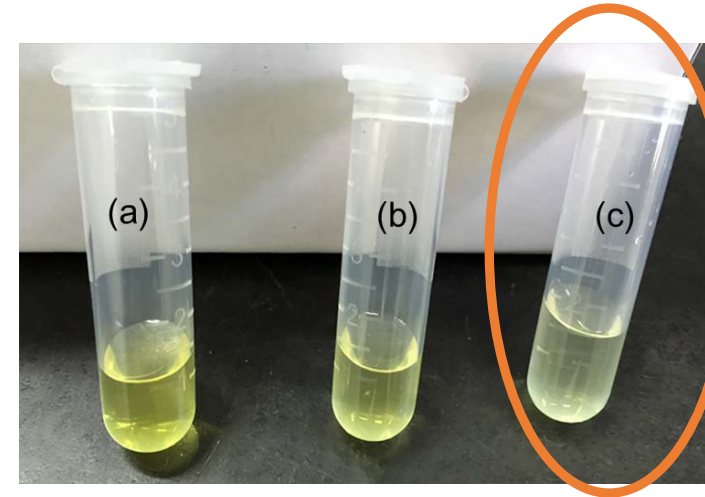
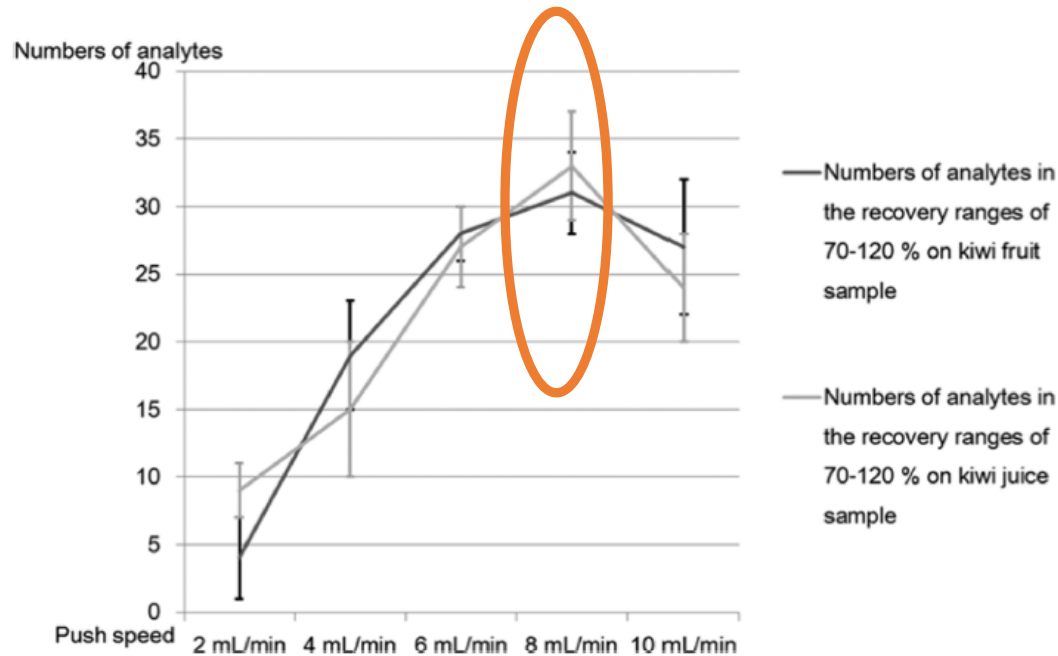


Method Editor

III. m-PFC automatic device and evaluation

——Semi-automatic m-PFC

- Optimization of m-PFC Cycles
- Optimization of automated m-PFC Volume
- Optimization of pushing speed
- Optimization of pulling speed

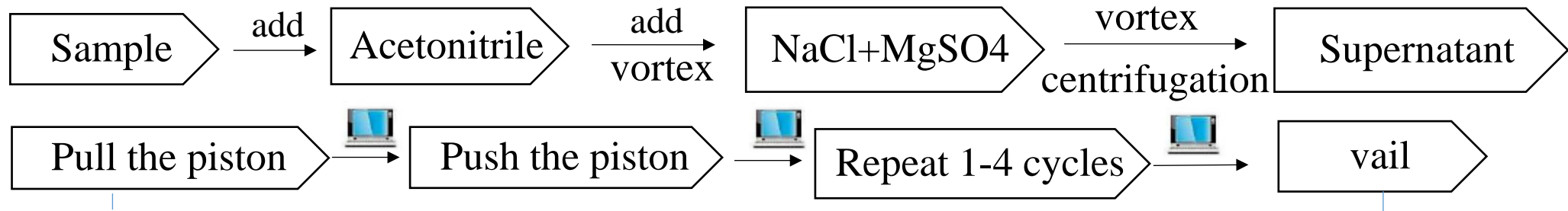
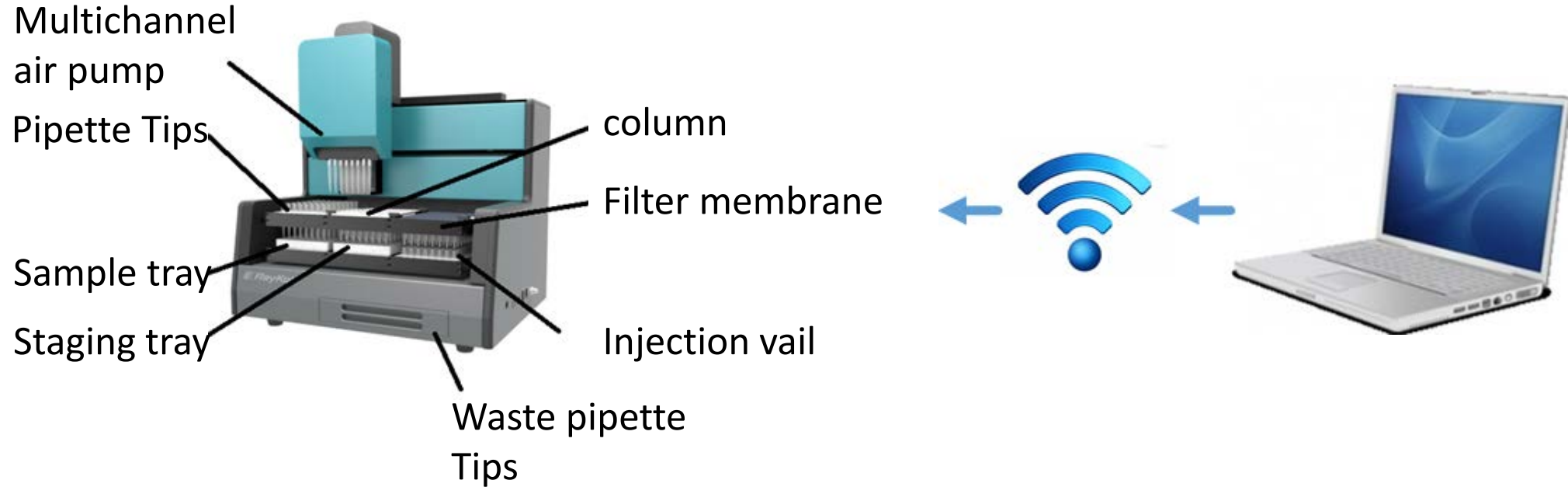


Comparison at different pushing speeds on the kiwi fruit sample spiked with the pesticides at 0.1 mg/kg:

(a) 4 mL/min; (b) 6 mL/min; (c) 8 mL/min.

III. m-PFC automatic device and evaluation

Multi-channel m-PFC /FC device





Comparison of the proposed method with mean recoveries and RSD in celery at 0.05 mg/kg

| m-PFC | automatic one-step m-PFC | | manual m-PFC | |
|---------------------|--------------------------|----------|------------------|----------|
| | Average recovery | RSD(n=3) | Average recovery | RSD(n=3) |
| methomyl | 96% | 1.1% | 92% | 2.5% |
| clothianidin | 98% | 2.5% | 97% | 2.9% |
| imidacloprid | 94% | 1.6% | 93% | 6.4% |
| acetamiprid | 95% | 1.0% | 94% | 2.8% |
| acephate | 86% | 2.4% | 84% | 2.7% |
| imazalil | 91% | 2.1% | 84% | 2.2% |
| pyrimethanil | 77% | 1.3% | 76% | 3.3% |
| thiophonate-methyl | 92% | 0.6% | 85% | 2.0% |
| flutriafol | 99% | 1.7% | 96% | 1.9% |
| metalaxyl | 101% | 1.8% | 96% | 2.9% |
| chlorantraniliprole | 88% | 2.2% | 87% | 2.3% |
| kresoxim-methyl | 101% | 1.5% | 94% | 2.7% |
| boscalid | 85% | 1.1% | 85% | 3.0% |
| tebuconazole | 97% | 1.7% | 91% | 1.5% |
| tebufenozide | 98% | 3.4% | 95% | 2.6% |
| difenoconazole | 98% | 0.7% | 96% | 2.5% |
| pyraclostrobin | 86% | 1.3% | 86% | 1.4% |
| trifloxystrobin | 93% | 1.7% | 91% | 3.2% |
| hexythiazox | 96% | 1.7% | 99% | 3.0% |
| pyridaben | 101% | 1.1% | 102% | 2.4% |

massive determination high efficiency

robust and reproducible

high-throughput

Automatic m-PFC process and filtration with filter membrane



Application of automatic multi-channel m-PFC device

Evaluations: 48 pesticide residues in green tea

82 pesticide residues in mulberry leaves and processed tea

Recovery ✓

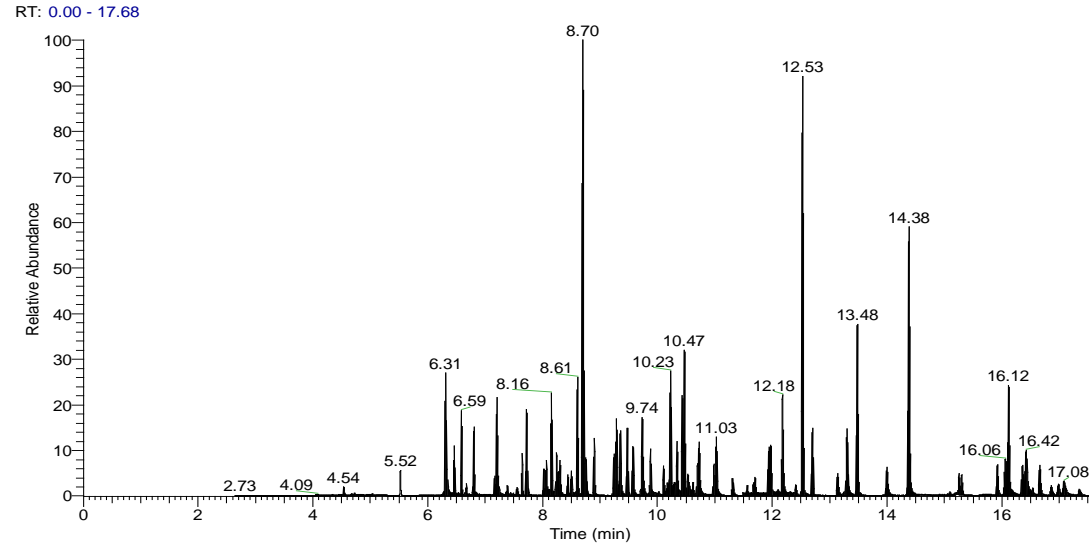
Matrix effect ✓

RSD ✓

LOD ✓

Linearity ✓

LOQ ✓

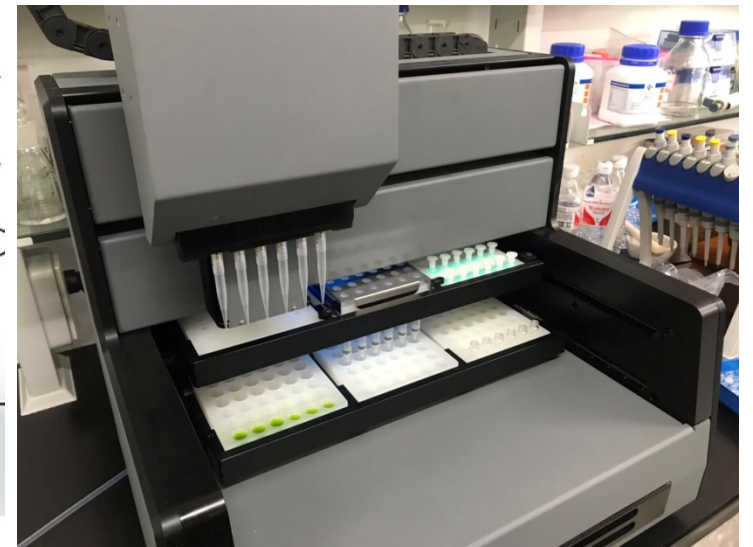
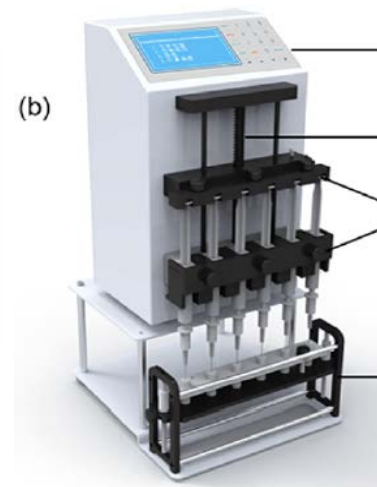
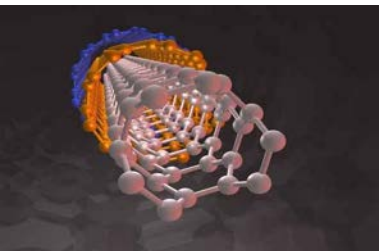


Comparison of the proposed method with other automatic m-PFC methods in mulberry processed tea

| Method | Recoveries (%) | RSD (%) | Sample number of batch process | Cleanup time cost per sample (min) | Automated filter |
|-------------------------|----------------|-------------|--------------------------------|------------------------------------|------------------|
| Automatic multi-channel | 73-116 | 2-13 | 48-64 | 0.8 | Yes |
| Automatic m-FC | 76-118 | 5-17 | 24 | 2-3 | No |
| Semi-automatic m-PFC | 72-124 | 1-19 | 6 | 2 | No |

Summary & Discussion

- 1. **MWCNTs, Nitrogen-enriched Carbon** : effective d-SPE materials
- 2. **m-PFC/FC**: rapid, potential high-throughput analysis
- 3. m-PFC **automatic** devices : speed, high efficiency, labor-saving , easy, robust and more reproducible





Thank you for your attention !

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